

Peer-review of the Wildlife Conservation Plan, prepared by the Wildlife Institute of India (WII) for the Etalin Hydropower Project, Dibang Valley, Arunachal Pradesh

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CONTRIBUTORS LISTED ALPHABETICALLY

Anindya Sinha, PhD, National Institute of Advanced Studies, Bengaluru
Anirban Datta Roy, PhD, Independent researcher
Arjun Kamdar, National Centre for Biological Sciences, Bengaluru
Aparajita Datta, PhD, Senior Scientist, Nature Conservation Foundation, Bengaluru
Chihi Umbrey, MSc, Department of Zoology, Rajiv Gandhi University, Itanagar, Arunachal Pradesh
Chintan Sheth, MSc, Independent researcher
M. Firoz Ahmed, PhD, Scientist F, Head, Herpetofauna Research and Conservation Division, Aaranyak, Guwahati
Jagdish Krishnaswamy, PhD, Convenor and Senior Fellow, Ashoka Trust for Research in Ecology and the Environment, Bengaluru
Jayanta Kumar Roy, PhD, Senior Researcher, Herpetofauna Research and Conservation Division, Aaranyak, Guwahati
Karthik Teegalapalli, PhD, Independent researcher
Khyanjeet Gogoi, TOSEHIM, Regional Orchids Germplasm Conservation and Propagation Centre, Assam Circle
Krishnapriya Tamma, PhD, Azim Premji University, Bengaluru
Manish Kumar, PhD, Fellow, Centre for Ecology Development and Research, Uttarakhand
Megha Rao, MSc, Nature Conservation Foundation, Bengaluru
Monsoonjyoti Gogoi, PhD, Scientist B, Bombay Natural History Society
Narayan Sharma, PhD, Assistant Professor, Cotton University, Guwahati
Neelesh Dahanukar, PhD, Scientist, Zoo Outreach Organization, Coimbatore
Rajeev Raghavan, PhD, South Asia Coordinator, South Asia IUCN Freshwater Fish Specialist Group
Rameshori Yumnam, PhD, Member, IUCN Freshwater Fish Specialist Group
Sanjay Molur, PhD, Zoo Outreach Organization, Coimbatore
Sayan Banerjee, MPhil, National Institute of Advanced Studies, Bengaluru
Shashank Dalvi, MSc, Independent researcher
Shantanu Joshi, MSc, National Centre for Biological Sciences, Bengaluru
Rohan Menzies, MSc, Nature Conservation Foundation, Bengaluru
Rohit Naniwadekar, PhD, Nature Conservation Foundation, Bengaluru
Umesh Srinivasan, PhD, Assistant Professor, Indian Institute of Science, Bengaluru

EXECUTIVE SUMMARY

A group of Indian scientists including botanists, entomologists, ornithologists, mammalogists, herpetologists, aquatic fauna specialists, hydrologists, geographers and social scientists, many with research experience in Arunachal Pradesh, including in the Dibang Valley, have conducted a peer-review of the Wildlife Institute of India's (WII) Technical Report titled 'Wildlife Conservation Plan for the impact zone of Etalin HEP, Dibang Valley District, Arunachal Pradesh' (hereafter, the Report). The Report was prepared in response to the Forest Advisory Committee's (FAC) recommendation to conduct "a multiple seasonal replicate study on biodiversity assessment of the catchment area" of the 3097 MW Etalin Hydro Electric Project (HEP) in Dibang Valley, Arunachal Pradesh. Overall, the review encountered considerable deficiencies, contradictory claims and scientific biases which have compromised the quality and the veracity of the Report's findings and conclusions. Below are some noteworthy findings:

1. The entire study was conducted in under 5 months from February to June 2018. This cannot be considered a 'multiple seasonal replicate' study and does not represent three seasons in Arunachal Pradesh with distinct rainfall and weather patterns: (a) October to February: relatively dry season/winter; (b) March-April: spring/pre-monsoon; and, (c) May to September: summer/monsoon.
2. **Flawed methodologies and lack of rigour:** The Report suffers from several technical shortcomings:
 - a. The sampling was done in a much smaller area ('Zone of Influence', ZoI) than will see the direct and indirect impacts of the HEP. Even within the ZoI, not all grids were surveyed, ignoring the potentially disastrous impact of the HEP on yet undiscovered and endemic taxa in a biodiversity hotspot. Most surveys were conducted along roads and other anthropogenically impacted areas using unscientific and biased methods. Not all elevations were sampled despite ample evidence that elevational gradients contribute to high biodiversity in the Eastern Himalayas.
 - b. Birds were surveyed outside the breeding season significantly reducing detection rates.
 - c. Several groups of taxa were not surveyed at all, including numerous insect orders such as Hymenoptera (bees, wasps and ants), Diptera (flies), Orthoptera (grasshoppers, locusts and crickets), and arthropod species such as scorpions, and pseudo-scorpions, and other taxa such as crustaceans (crabs), molluscs (snails), and protozoans despite evidence of high levels of diversity and endemism in the Dibang River basin.
 - d. Approaches and methods used to analyse field data and produce results are inadequate, not clearly explained and, often, not scientifically recognised. Importantly, species accumulation curves, a key method to analyse and report data on species richness and diversity, have not been prepared for most taxa. Fish is the only taxa for which accumulation curves have been prepared but they clearly show inadequate sampling. Furthermore, nowhere are the links between topography, habitat type, biodiversity presence, local people's priorities and potential HEP impacts clearly established.
 - e. The Report does not refer to recent and relevant peer-reviewed work on social, ecological, physical and geomorphological aspects of the study region, including one by some of the Report's own authors that found tigers within 10 km of the HEP site. Instead, the Report relies on limited, often outdated material.
3. **Flawed inference:** Because of serious methodological and analytical deficiencies and exclusion of highly diverse taxa such as insects and arthropods, the Report underestimates the study area's biodiversity values and the HEP's impacts, particularly on range-restricted, threatened and

endemic species. Comparisons with published research from Dibang Valley shows that the Report under-reports hundreds of species of butterflies and other groups of insects, and birds, and tens of species of orchids, mammals and herpetofauna. The report's species checklists contain repetitions, lack of proper taxonomic classification and wrong distributions including 12 species of butterflies that are not known to occur in Northeast India at all.

4. ***Dibang is exceptionally biodiverse:*** Even with the short survey conducted using biased sampling methods, the Report provides direct evidence of 230 bird, 159 butterfly, 112 spider, 51 moth, 31 reptile, 14 amphibian and 21 mammal species. Amongst these are several endemic (e.g. seven species of birds), range-restricted (e.g. six bird and three butterfly species) and threatened (e.g. eight mammal species) species, many of which are included in Schedule I of the Indian Wild Life (Protection) Act, 1972.
5. Despite numerous admissions that highlight the species richness and diversity of the region, the Report ignores its own findings to outline mitigation measures for some taxa while observing that “it was not possible to suggest any threatened species and habitat specific conservation plan” for others (e.g. mammals). Firstly, it is not clear on what basis the FAC's singular mandate of *'biodiversity assessment'* was converted into a Wildlife Conservation Plan. Secondly, the few mitigatory measures recommended for some specific faunal groups in the form of butterfly, reptile parks and nest boxes cannot be considered as well-designed ecologically meaningful measures to compensate for the local entire ecosystems that provide vital environmental and societal benefits. Overall, this assumes the project as *fait accompli* implying that the Report's findings have no bearing on the FAC's decision to approve the project, ultimately making this exercise appear futile.
6. There appears to be a minimization of the negative impacts of the HEP and local concerns about them throughout the Report. The section that relate to assessing socio-cultural impacts of the HEP and suggest mitigatory measure assume a patronizing attitude towards the local Idu Mishmi people and emerge from a lack of a nuanced understanding of socio-cultural dynamics and interdependencies between people and the natural environment. The Report recommends large-scale agriculture and a shift to cash crops, without considering the impact of such large scale (possibly monoculture) cultivation on biodiversity, local livelihoods and well-being. Some of the job opportunities listed such as welder, fitter, plumber, electrician etc. are largely alien to most highland farmers and rests on the assumption that people can make an effortless transition to new livelihoods immediately after the shock of resettlement.

Crucially, studies that inform high-level decision-making on historically significant projects, such as the Etalin HEP (which would be one the largest hydropower projects in the country), must go through a transparent and scientifically recognised peer-reviewed process given the pitfalls, numerous discrepancies and gaps highlighted in this review. Such decisions have irreversible impacts on lives, livelihoods and the environment.

INTRODUCTION AND BACKGROUND

On 28 February 2017, the Forest Advisory Committee (hereafter FAC) met to discuss the 3097 MW Etalin Hydropower Project (hereafter HEP, or ‘the project’) to be developed by Jindal Power Limited (hereafter User Agency) in Dibang Valley district. In the minutes that were released later (F.NO. 8-20/2014-FC), the FAC found the Environmental Impact Assessment (EIA) submitted by the User Agency “inadequate” and recommended that a “*multiple seasonal replicate study on biodiversity assessment of the catchment area*” of the 3097 MW Etalin HEP in Dibang Valley district be conducted by “an internationally credible institute”. The Wildlife Institute of India (hereafter WII), Dehradun was chosen to conduct the said study (vide letter no. FOR-279/CONS/2010/Vol-I/ 836-40, 23rd June 2017 from APCCF and Nodal Officer (FCA), Arunachal Pradesh). In 2019, the WII produced an extensive Technical Report (TR No/2019/01, hereafter ‘the Report’) titled ‘Wildlife Conservation Plan for the impact zone of Etalin HEP, Dibang Valley District, Arunachal Pradesh’. The Report assesses the status of various taxonomic groups including mammals, avifauna, entomofauna, herpetofauna and flora in the HEP site. It also documents the biodiversity value for and the natural resource dependence of the local Idu Mishmi people in the project site. Finally, it evaluates the impacts of the proposed project on the aforementioned taxa and natural resource needs of the local people and drafts a mitigation and conservation plan along with a financial budget for its implementation.

A group of Indian scientists, including botanists, entomologists, ornithologists, mammalogists, herpetologists, aquatic fauna specialists, geographers, hydrologists, and social scientists, who have multiple years of research experience in Arunachal Pradesh, including in Dibang Valley, have conducted a peer review of the Report. Overall, the review encountered considerable deficiencies and scientific biases in the Report which have compromised the quality and the veracity of its findings and conclusions. Before proceeding to an in-depth taxon-wise review of the Report, below are some key general observations:

- (1) The FAC recommended a “*multiple seasonal replicate study on biodiversity assessment of the catchment area*”. The entire study however appears to have been conducted by the WII over a short period from February 2018 to June 2018, which is under 5 months and cannot be considered a ‘multiple seasonal replicate’ study. February and March have been taken as winter/pre-monsoon and April to June as summer/monsoon. These do not represent seasonal patterns in Arunachal which has at least three seasons with distinct rainfall and weather regimes: (a) October to February: relatively dry season/winter; (b) March-April: spring/pre-monsoon; and, (c) May to September: summer/monsoon.
- (2) The Report is a Wildlife Conservation Plan with the final chapter (Chapter 7) dedicated to mitigation and conservation measures. It is not clear on what basis the FAC’s singular mandate of ‘*biodiversity assessment*’ was converted into a Wildlife Conservation Plan.
- (3) The Report does not refer to recent and relevant peer-reviewed work on social, ecological, physical and geomorphological aspects of the study region, instead relying on limited and often outdated material. The sections below highlight some of the crucial literature that should have been perused.
- (4) Throughout, but particularly in Chapters 6 and 7, the Report segregates the impacts of the project neatly between ‘Physical’, ‘Biological’ and ‘Social’ components. Such a categorization represents a highly narrow, misinformed and flawed understanding of the interconnections between physical, biological and social processes. In developing this schema, where the assumption is that the construction of many components of the HEP will only have biological but no knock-on social impacts (see impact matrix in Table 6.1), the Report seems to have entirely

ignored vast and widely-popular multi-decadal literature on the interconnections between social and ecological systems (SES) (e.g. Adger ,2000; Young et al., 2006). If changes in ecology indeed have no knock-on impacts on people’s social lives, then how does the Report envisage explaining the devastating social, cultural and economic impacts of decidedly natural/ecological phenomena such as climate change, locust infestations, and zoonotic diseases such as the ongoing COVID-19, to name a few.

- (5) Even in the short survey conducted using biased sampling methods within a limited a study area, the Report provides clear evidence for the existence of rich biodiversity. While many more species previously recorded from the study area have been omitted (highlighted in taxa-specific sections), it nonetheless collected direct evidence of 230 bird, 159 butterfly, 112 spider, 51 moth, 31 reptile, 14 amphibian and 21 mammal species. Amongst these, they found several species that are endemic (e.g. seven species of birds), range-restricted (e.g. six bird and three butterfly species) and threatened (e.g. eight mammal species), many of which are included in Schedule I of the Indian Wild Life (Protection) Act, 1972 (IWPA), affording them the highest degree of protection. The Report makes numerous statements that highlight the species richness and diversity of the region repeatedly stressing that it is critical to preserve these sites (e.g., “The presence of Rare, Endangered or Threatened [RET] or species of conservation significance along both the rivers, shows the importance of the habitat and plant species at each project activity site/impact zone for these species. Disturbance of any sort will lead to disappearance of that species...” Page 82). Yet, and ironically, it deliberately ignores these findings to outline mitigation measures for some taxa (e.g., butterfly parks, nest boxes etc.) while observing that “it was not possible to suggest any threatened species and habitat specific conservation plan” for others (e.g. mammals).
- (6) At various points, the Report states that species and habitat specific conservation plans are not possible due to species diversity and their diverse dietary and foraging patterns, yet recommendations are still made for butterfly parks, reptile parks, and habitat restoration that clearly will not replace the loss of natural habitat and address the direct impacts on species. While the appropriateness and viability of these mitigative measures are assessed in greater detail in the taxon-specific sections, this process assumes the project as *fait accompli* implying that the Report’s findings have no bearing on the FAC’s decision to approve the project, ultimately making this exercise appear futile.

The main text of this review is divided into nine sections, each corresponding to a specific focal area of the Report. It begins with a critique of the criteria used to delineate the HEP’s area of impact (‘Zone of Influence’) and the analytical framework of the Report. Section 2 evaluates the geospatial analyses used to estimate vegetation types. This is followed by seven sections, each presenting a thorough review of field data collection and analytical methodology, suitability and accuracy of the findings and validity of the conclusions for the specific taxon assessed in the Report. While a detailed social science critique of the methodology and results of the socio-cultural surveys is outside the scope of this review, the last section offers a broad response to the Report’s findings on socio-cultural value of biodiversity. The review concludes with critical reflections on the reliability of the Report’s findings to inform decision-making given the issues identified in prior sections.

Large projects such as the Etalin HEP are multifaceted issues that require attention to many different, yet interrelated, aspects including, but not limited to, socio-cultural realities, political and economic viability, engineering design, the natural environment, and local needs and perspectives. Decision-making on such historically significant projects is, therefore, unarguably complex. This review

underscores the importance of paying attention to scientific processes, findings and realities, which should be integral to informing any development of this scale.

SECTION 1: REVIEW OF ZONE OF INFLUENCE AND ANALYTICAL FRAMEWORK

A detailed analysis of the process of delineation of the study area (the ‘Zone of Influence’) for biodiversity assessment, field sampling methodology and data analysis framework applied across all floral-faunal groups (Chapters 4 and 5) raises serious doubts over the reliability of the Report’s findings on biodiversity richness and the HEP’s expected impacts. Furthermore, nowhere are the links between topography, habitat, biodiversity, local people’s priorities and potential impacts clearly established. These links are critical to ascertain which habitats and community land-use will be at high risk from landslides or erosion due to project related activities and which of these high-risk areas are occupied by endangered flora and fauna. Thus, the lack of a robust holistic scientific framework risks downplaying the threats to the region’s ecology, hydrology and people because of the HEP. Specific comments on the overall lack of scientific rigour in the Report are as follows:

Impacts of HEP extend beyond the Zone of Influence

The Report broadly defines the ‘Zone of Influence’ (hereafter ZoI) as the farthest influence of the HEP (Page 35). However, this delineated zone of 112 grids (1×1 km each) does not consider areas upstream and downstream of the dams that will be severely altered due to storage of sediments and changes in flow regimes. Neither does it include the environmental impacts of subsurface tunnel construction. Additionally, Dibang valley lies in the zone-V of the earthquake hazard zone making it highly prone to earthquakes and its associated affect like landslides. The report undermines the severity of risks entailed in carrying out massive infrastructural projects in fragile landscapes and presents a myopic perspective that the HEP’s impacts on biodiversity are unrelated to human well-being.

Even within the limited ZoI, the Biodiversity Conservation Plan does not account for the damage to the greater landscape by construction of over 50 km of new roads and widening of an additional 30 km of existing roads. The true ZoI due to road construction, quarrying and debris dumping is likely to be much larger because of the extensive slopes on either side of the steep river valley, and their very-high susceptibility to landslides. The landslide susceptibility of the region was not considered despite a global landslide susceptibility map developed by NASA, available at no charge (Stanley and Kirschbaum, 2017) (Page 183). The Report fails to appreciate previous studies globally and in Dibang Valley on the detrimental effects of landslides on forests, rivers, people, and biodiversity (Sassa and Canuti, 2009; Athreya and Sheth, 2016). The areas proposed for land acquisition have “high” and “very high” susceptibility to landslides according to the said global map (Stanley and Kirschbaum, 2017). This is evident from satellite images acquired before and after 2018 showing several ongoing landslides due to road widening and recent extreme precipitation. In addition, effective management of debris is extremely important as waste from an affected site may end up in an unaffected site thus extending the ZoI. Therefore, the ZoI appears greatly under-defined and not based on a careful consideration of topographical, hydrological, anthropogenic and geological factors known to impact mountain ecosystems.

Inadequate sampling, yet Dibang Valley is rich

The overall biodiversity of the area is several orders of magnitude larger than has been reported (see below) since the study does not appear to have taken microhabitats into account. A grid size of 1×1 km

may not be appropriate for all taxa and the Report gives no justification for such a study design. Smaller taxa need to be sampled for richness at a much finer spatial scale that adequately samples all habitats, micro-habitats, elevations and stream orders. A nested survey design may be best to survey multiple taxa.

Species accumulation curves are standard practice in biodiversity assessments as they provide an estimate of the total species richness as a function of area and time, and indicate the adequacy of a survey in representing the fauna of a particular area. Apart from species accumulation curves for fish, the Report does not present these curves for most other taxa (Page 62). The study also focuses on largely diurnal species (except for mammals). The sampled grids for all taxa (approximately 17 grids for plants, 26 for fish, 32 for mammals, 43 for entomofauna, 59 for birds were sampled out of 112) were restricted to areas that are accessible (along Etalin-Anini and Etalin-Maliney road), disturbed and designated for land acquisition. Given Dibang's challenging mountainous terrain, sampling may not be possible everywhere; however, this drawback was not accounted for statistically or acknowledged in the report. Thus, species richness is expected to be high for sampled grids and low for unsampled grids (Page 53). Despite these statistical flaws and deficiencies in sampling strategy, coupled with a less-than-ideal sampling season, the Report finds endemic and RET species even in ecologically disturbed areas such as roads, contradicting its claim that the impact potential in undisturbed areas would be low (Map 6.1; Page 148).

Importantly, within the ZoI, the grids that were not surveyed were assigned no biodiversity values. Yet, the potential impact of the HEP on those grids was ascertained to be "low" (Page 148). This approach is not based on any of the scientifically published methods that are used to score the impact on biodiversity due to HEPs.

Mid-elevations will be richest in Dibang Valley

Species richness peaks at different elevations for different taxa. These are well established ecological rules (Colwell and Lees, 2000; McCain and Grytnes, 2001). However, the study does not sample across different elevations to ascertain species richness-elevation relationships. Prior work in Western Arunachal demonstrates that species richness increases with elevation in certain taxa (Marathe et al., 2020; Sheth 2020 pers. comm.). Similarly, Roy et al. (2018) surveyed an elevation range from 200–3500 m in Dibang river basin documenting 38 amphibian species, 36 of which were found between 800–1500 m, the elevation range where the proposed HEP and associated infrastructure will be located. Long-term amphibian studies in Western Arunachal also show this mid-elevation zone to contain the highest number of species (Athreya and Sheth, 2016). Further, the mid-elevational peak in species richness observed in the Eastern Himalaya also suggests that these regions act as important biogeographic transition zones, where taxa from different biogeographic regions overlap (Kreft and Jetz, 2013). The lack of sampling across multiple elevations in the ZoI may have likely underestimated the biodiversity value, evolutionary importance, and impact assessment of the HEP in the study area.

SECTION 2: REVIEW OF GEOSPATIAL DATABASE

The geo-spatial database has several methodological shortcomings with respect to the quality of satellite imagery and the image classification process. More importantly, the land-use land-cover data have not been integrated with datasets on biodiversity richness, geohazards and topography to

understand the importance of landscape drivers in sustaining high-levels of biodiversity in the region. This requires longer data collection over larger region to develop empirical multi-level models capable of comprehensively assessing the true extent of impacts of the proposed HEP. Specific issues with geo-spatial analyses in the Report are detailed below:

Satellite image analysis and interpretation

- While the Report does not explicitly state how many bands are used for land-use land-cover classification, it appears that only two bands (out of 9 medium-resolution bands) of Sentinel-2 and two bands (out of 9 medium-resolution bands) of Landsat 8 are used (Page 63). This is important as the image-classification accuracy tends to improve as more bands are used (Forkuor et al., 2018). Further, the images are only from the post-monsoon season when the sun-angle is low creating larger shadows obscuring many slopes (Cingolani et al., 2004). These shadows reduce the effective area that can be classified.
- The Report does not clarify how different land-use and land-cover categories are defined (e.g. what was the basis of defining a particular habitat as evergreen forest). Distinguishing between evergreen and/or secondary forest is difficult in Arunachal and even more so in this season especially without a robust ground-truth sampling design. While the Report states that ground-truthing data were collected, no such data, including the number of pixels for each land cover category, are presented for scrutiny (Page 60).
- The NDVI/MSAVI indices used in the Report are not ideal for image classification when used on their own as they simply provide an index of vegetative biomass (Jackson and Huete, 1991). For areas with high vegetation biomass, the more sensitive EVI (Enhanced Vegetation Index) is recommended. Further, the recommended best practices for land-use and land-cover classification are to provide a statistical measure of the performance of classification models (Olofsson et al., 2014). However, the Report does not provide any details or statistics (e.g. confusion matrix, commission-omission errors) to allow the accuracy of the classified imagery to be assessed.
- Due to its narrow-minded focus on the individual components of the biodiversity, the report fails to look at the impact of the proposed activities on the ecosystem processes that sustain the rich biodiversity. It also ignores the potential impact of climate change on ecosystem processes and biodiversity in the region. It misses the opportunity to utilise various freely available gridded climatic data products (precipitation, temperature etc.) to further explore the nature of interlinkages between climate and biodiversity in the region and how such a mega-project will exacerbate the potential impacts of warming, precipitation changes, phenological shifts and increase of extreme events on the flora and fauna of the region.

SECTION 3: REVIEW OF FLORA

Despite limited long-term botanical research in Arunachal Pradesh, the state is known to host an outstanding floral diversity (Rao and Hajra, 1986). Dibang Valley hosts several endemic plants, many of which are new to science and have been recorded from within the ZoI. Following are some noteworthy plants recorded in Dibang Valley:

1. New species of *Impatiens* (commonly known as Balsams) such as *Impatiens ashihoi*, *Impatiens albopetala*, *Impatiens dibangensis* and others. (Gogoi and Borah, 2015; Gogoi and Borah, 2016)
2. Araceae members (Aroids) such as *Arisaema gracilentum*, *Colocasia dibangensis*, and several others have been described as new to science, distribution and status of which are yet to be studied properly (Gogoi and Borah, 2013; Bruggeman, 2016).
3. *Sapria himalayana*, called ‘a floral wonder’, has also been recorded from Dibang Valley (Hohl and Sebastian, 2014).
4. Members of the family Gesneriaceae are found in great diversity in Dibang Valley including many with a high ornamental value. The Report mentions *Henckelia mishmiensis* (*Chirita mishmiensis*) which is a Dibang Valley endemic that only grows in a specific niche.

Incomplete documentation of floral wealth

A mere 1.19 % of the ZoI was sampled for its floral diversity. The ZoI has 112 grids (1x1 km), of which 133 vegetation plots of 10 x 10 m were sampled. Yet, even with such limited sampling, the Report mentions that 398 plant species belonging to 106 families and 286 genera were encountered indicating very high floral diversity. However, even this is likely to be a significant underestimate based on the findings of previous studies (e.g. Liden and Adhikari, 2019) that have reported numerous new records for India from the high elevation areas of Dibang Valley, postulating that a similar number of new reports are expected from mid-to-low elevation areas. In particular, the tree diversity estimate mentioned in the Report seems low when compared to tropical and subtropical forests from other studies in neighbouring areas within the state (Borah and Garkoti, 2011; Dutta and Devi, 2013a,b; Sarkar and Devi, 2014; Sarma and Borah, 2014; Borah et al., 2016; Saikia and Khan, 2016; Bora and Bhattacharyya, 2017; Borogayary et al., 2017; Barua et al., 2018). Since Dibang Valley is a hotspot for discovery of new plant species, the herbarium species collected during fieldwork should urgently be re-examined (if they have been retained) as many of these unidentified specimens could turn out to be new to science. Much of this identified and yet-to-be-identified botanical diversity is at risk from the proposed HEP and the Report does not do an adequate job of identifying these potential impacts.

Endemic orchid diversity not adequately sampled

The Report similarly underestimates orchid diversity identifying only 35 species for the ZoI's. All of the recorded orchids are extremely common species and a comprehensive survey with taxonomic expertise will reveal more species, including those that are rare and endemic. Existing studies have already documented 117 species of orchids belonging to 44 genera from Dibang Valley. Among these, 86 species are epiphytes, three are epiphytic as well as terrestrial, and 31 species are terrestrial including four saprophytes (Bhaumik and Pathak, 2010). More recent surveys have reported up to 200 species with the highest diversity found in low-to-mid elevations areas similar to the ZoI (Gogoi 2020 pers. comm.).

Under-reporting ethnomedicinal knowledge

The Report records only nine species of medicinal plants used in the area (Table 5.54). This is difficult to believe as prior research has established that the Idu Mishmi have a vast knowledge of medicinal plants and use them regularly for a variety of ailments. Haridasan et al. (1995), reported more than 500 species of medicinal plants from Arunachal Pradesh. Although ethnobotanical studies have been scarce

in the Dibang Valley, recent work has found 36 species in Lower Dibang Valley (Tangjan et al., 2011), more than 80 species between the two Dibang districts (Shankar and Rawat, 2008) and 55 species within the Dihang Dibang Biosphere Reserve (Ghosh et al., 2014).

SECTION 4: REVIEW OF ENTOMOFAUNA

Recent reports of decline in insect populations worldwide are alarming (Hallmann et al., 2017; Leather 2018; Sánchez-Bayo and Wyckhuys, 2019), not least because insects provide crucial pollination services. Further, 73% of the world's cultivated crops are pollinated by honeybees (Raj et al., 2012). Arunachal Pradesh supports a rich diversity of bees with about 49 species recorded to date in limited surveys. These include the family Megachilidae, genus *Ceratina* and 13 other bee species which were recently recorded for the first time (Saini et al., 2018). Very limited work exists on the honeybees of Dibang Valley and no comprehensive research has been done on the numerous other wild bee species of the district. So far only *Apis laboriosa*, *Apis dorsata* and *Apis cerana* have been recorded from Dibang Valley (Gogoi et al., 2018).

Members of the order Lepidoptera, which includes butterflies and moths, are another diverse group of significant pollinators, second only to bees. The Lepidoptera, too, have experienced sharp population declines (Fox 2013; Langevelde et al., 2018; Thogmartin et al., 2017). Beyond being a key pollinator, this insect group is also a significant bio-indicator of climate change, deforestation and habitat degradation. Lepidopterans are also prey for birds, mammals and reptiles, and their larvae are mostly plant herbivores. Finally, due to a paucity of long-term research across Arunachal Pradesh, experts believe that numerous butterfly and moth species new to science are yet to be described from this landscape.

Dibang Valley as an important area for endemic and rare entomofauna

Dibang Valley is believed to have more than 500 species of butterflies. Many subspecies of butterflies occurring in S.E. Tibet and Yunnan also occur up to Dibang Valley (for example, the nominotypical subspecies of Chocolate Tiger *Danaus melaneus* butterfly is distributed in S.E. Tibet, Yunnan and Dibang Valley). The Brahmaputra river basin has been a barrier to the dispersal of many butterfly species resulting in high rates of endemism and speciation in Dibang Valley. For example, the Dibang Valley endemic Roy's Argus *Callerebia dibangensis* was only described in 2013 while many hairstreaks, rings, skipper butterflies etc in the region are yet to be described. This rich diversity of butterflies and their colourful unique patterns (like the Northern Jungle Queen) have found a place in Mishmi lives and some of their traditional weaving patterns are inspired from butterflies (Elwin 1959). Some other range-restricted butterflies of Dibang Valley include False Tibetan Cupid *Tongeia pseudozuthus*, Chinese Silverline *Spindasis zhengweilie*, Khaki Silverline *Spindasis rukmini*, Evans Silverline *Spindasis evansii*, Tiger-mimic Admiral *Limenitis rileyi*, Mottled Argus *Callerebia narasingha*, Tibetan Brimstone *Gonepteryx amintha thibetana*, Grey Commodore *Bhagadatta austenia purpurascens*, Abor Freak *Calinaga aborica* (see Appendix I for an updated butterfly checklist for Dibang Valley). All of these species, including endemics such as *Callerebia dibangensis*, occur in the elevational range of Etalin HEP raising serious concerns of the project's impact on their survival. Concerns and deficiencies in the entomofaunal assessment of the Report are detailed below with a specific emphasis on Lepidopterans.

Threatened and endemic species missed in ZoI

The report's account of (159 species of butterflies, 11 species of odonates and 51 species of moths) is low and may not be a true representation of the study area's species richness. There are many threatened and endemic entomofauna even in the ZoI that have not been reported. Further, many species mentioned in the Report have been erroneously assigned to different taxonomic categories. Despite this, recording 200+ species of entomofauna within a limited sampling period and effort underscores the immense evolutionary and ecological wealth of this area. Our specific comments are as follows:

- **Butterflies:** 354 species of butterflies have been reported from Dibang Valley in the last 10 years (Gogoi 2020, unpublished data.), while up to 500 species are believed to exist in the region. However, the Report mentions a mere 159 species from the project site. The riparian habitat within the ZoI is likely to have around 290-300 butterfly species based: (1) a 4-month survey in a similar habitat in Lower Dibang Valley that recorded 294 species (Gogoi 2012); (2) surveys in comparable habitats in other parts of Arunachal Pradesh where up to 700 species have been recorded (Sanjay Sondhi, pers. comm, unpublished report).
- **Macro-invertebrates:** Benthic macro-invertebrates are considered one of the most important bioindicator groups for freshwater ecosystems. The Report poorly studies these key taxa identifying up to family level only. Further, each family is considered a different taxon (Tables 5.48 and 5.49). This is a gross underrepresentation of the actual diversity as each family contains several species. Using these family-level as opposed to species-level data, the richness of EPT taxa (Ephemeroptera, Plecoptera, Tricoptera) has been plotted in Figures 5.1 and 5.2, erroneously obtaining a very low number of 7-8 in all sites. This is misleading and an inaccurate representation of actual species richness at these sites.
- **Odonata:** The Report states that eleven odonate species occur in the ZoI's riparian habitat. This is a shockingly low number given that other studies, even from urban areas have reported many more species. More than 60 species are expected to be found in the Etalin area with adequate sampling and correct identification (Arajush Payra 2020 pers. comm.). The Report records only two damselflies, which is a serious underestimate for Dibang Valley. Even common species observed at virtually all water bodies across India, including across Dibang Valley such as *Ischnura rubilio*, *Ceriagrion coromandelianum*, *Calicnemia miles* and *Pseudagrion rubriceps*, have not been included, indicating inadequate sampling. A species new to India, *Echo perornata* recently reported from Hunli-Anini road in Dibang Valley (Gogoi and Payra 2019), does not make it into the Report as well. As for dragonflies, eight families are known to occur in India, all of which are found in Arunachal Pradesh as well (Subramanian and Babu, 2017). However, only the most common family Libellulidae is reported. The dragonfly family Gomphidae and damselfly family Lestidae are incorrectly reported with benthic invertebrates (Page 123-124) and not with the odonates species list (Page 255).
- **Moths:** The report's account of 51 moth species is an extremely low number for this group. Around 10,000 species of moths occur in India and the Etalin area is likely to have up to 600-700 species. In other similar habitats, such as in Eaglenest and Talle Valley wildlife sanctuaries (WLS), the list of identified moth species exceeds 400 (Sanjay Sondhi, pers. comm, unpublished report), with at least a similar number of additional species that are believed to be recorded if long term surveys are conducted. Many species in this habitat type and elevation are yet to be discovered. For example, in 2017, the Apatani Glory *Elcysma ziroensis*, a species new to science was described from Tale Valley WLS. It remains the only known location in the world that this species is known from. Similar moth diversity is expected from the project site.

The Report has entirely ignored the evaluation of butterflies and other entomofauna using their criteria of 'Rare Endangered and other Threatened' species (RET). For example, Himalayan Mottled Argus *Callerebia narasingha narasingha* (Moore, 1857), included in the report's butterfly checklist, is endemic to Eastern Himalaya and would qualify as an RET species. Similarly, a Dibang Valley endemic - Roy's Argus *Callerebia dibangensis* - likely to occur in Etalin given the elevation, has not been assessed.

The review of secondary literature is rather inadequate and important references have been left out. Some notable published omissions include several populations of range-restricted butterflies such as the False Tibetan Cupid *Tongeia pseudozethus* (Huang, 2001), known to occur between Hunli-Anini; Tibetan Brimstone *Gonepteryx amintha thibetana* reported from Dri valley and Mayodia-Anini road (Sondhi and Roy, 2013; Das and Gogoi 2020 pers. comm.); and the Blue Posy *Drupadia scaeva cyara*, which was recorded for the first time in India from the area around Etalin (Das et. al., 2018). None of these important references have been included effectively downplaying the potential impacts of the HEP to these extremely rare and endemic species [Page 194].

Ineffective mitigation plans

The Report has indicated that four to five 'Open Butterfly Parks' will be set up as part of a Species Group Conservation Plan to attract a portion of the 159 species of butterflies identified. However, the host plants of these threatened and endemic butterflies are still unknown, questioning the viability of such measures. Baseline data on larval host plants for many of Dibang's endemic and range-restricted species, such as *Callerebia dibangensis*, are still missing. The purpose of a butterfly park will be defeated if the only known habitat and host plants for these endemics are lost. In any case, the establishment of a butterfly park cannot compensate for the direct loss of the habitat/microhabitats of butterflies and the consequent impacts on their population, persistence and survival. It also does not suggest any mitigation for any other entomofaunal groups that were assessed and will be affected.

Additional errors in entomofauna sections

There were also several errors in the report, 12 species of butterflies mentioned in the checklist are not distributed in Northeast India at all. Most of these species either occur in the Western Himalayas or Peninsular India. Some other species in the checklist are doubtful as their presence in Arunachal Pradesh is unlikely and has not been confirmed. Further, the butterfly checklist has numerous errors in assigning correct family names, while one species has been repeated twice with different generic combinations. If the researchers had consulted existing literature, a more accurate account could have emerged. These details are mentioned below:

- The extremely rare Scarce Jester *Symbrenthia silana* is mentioned in the Report as Scarce Jester 'Symbrenthia silana de' [Page 172]. Firstly, the 'de' should have been 'de Niceville'. Secondly, the species is endemic to Eastern Himalaya, which has not been highlighted, which would make it an RET species.
- *Faulty distribution*: A total of 12 species of butterflies mentioned in the report's checklist do not occur in Northeast India at all. Most are distributed in the Western Himalayas or Peninsular India (Gasse, 2013). These species are:
 1. *Heliophorus oda* (Hewitson, 1865) Eastern Blue Sapphire
 2. *Curetis thetis* (Drury, 1773) Indian Sunbeam
 3. *Lycaena phlaeas* Small Copper
 4. *Euploea crameri nicevillei* Spotted Black Crow

5. *Euploea tulliolus* Dwarf Crow
 6. *Ypthima asterope mahratta* (Moore, 1884) Common Threering
 7. *Ypthima sakra sakra* (Moore, 1857) Himalayan Fivering (*parasakra* occurs in NE India)
 8. *Dodona durga durga* (Kollar, 1844) Common Punch
 9. *Papilio crino* (Fabricius, 1793) Common Banded Peacock
 10. *Pareronia hippia* (Fabricius), Common Wanderer
 11. *Pareronia* sp. Dark Wanderer
 12. *Pieris rapae meleager* (Hemming, 1934) Small Cabbage White
- The following species reported as occurring in Dibang Valley are doubtful:
 1. *Taractrocera maevius* Common Grass Dart
 2. *Graphium megarus megarus* (Westwood, 1844) Assam Spotted Zebra
 3. *Gonepteryx rhamni nepalensis* (Doubleday, 1847) Himalayan Brimstone
 4. *Charaxes moori* (Distant, 1883) Malayan Nawab
 - The following species are wrongly placed under Family Papilionidae; they should be under Family Pieridae
 1. *Appias lalage lalage* (Doubleday, 1842) Spot Puffin
 2. *Colias fieldii fieldii* (Menetries, 1855) Dark Clouded Yellow
 3. *Gandaca harina assamica* (Moore, 1906) Tree Yellow
 4. *Gonepteryx rhamni nepalensis* (Doubleday, 1847) Himalayan Brimstone
 - The following species are wrongly placed under Family Pieridae; they should be under Family Riodinidae
 1. *Abisara neophron neophron* (Hewitson, 1861) Tailed Judy
 2. *Dodona adonira adonira* (Hewitson, 1865) Striped Punch
 3. *Dodona dipoea dipoea* (Hewitson, 1865) Lesser Punch
 - The following species are wrongly placed under Family Nymphalidae; they should be under Family Papilionidae
 1. *Papilio helenus helenus* (Linnaeus, 1758) Red Helen
 2. *Papilio paris paris* (Linnaeus, 1758) Paris Peacock
 3. *Papilio polytes romulus* (Cramer, 1775) Common Mormon
 - Two species mentioned are synonyms of each other and should not be listed separately. *Precis iphita* is a junior synonym of *Junonia iphita*
 1. *Junonia iphita iphita* (Cramer, 1779) Chocolate Pansy
 2. *Precis iphita iphita* Chocolate Soldier
 - Two out of the four photographs of dragonflies are wrongly identified (Page 91).
 1. The photo labelled as *Orthetrum taeniolatum* is an *Orthetrum luzonicum* immature male.
 2. The photo labelled as *Pantala flavescens*, one of the most common species of dragonflies worldwide, is a female *Orthetrum pruinosum*.
 - *Calicnemiinae* reported as the family for the two species below is actually a subfamily. The correct family name is *Platycnemididae* (Subramanian and Babu, 2017).
 1. *Calicnemia miniata* (Selys, 1886)
 2. *Calicnemia* sp. 2

In conclusion, the Report appears to consider the importance of butterflies (and other insects) only in terms of pollination services (“it is very important to conserve butterfly species, as they help in pollination” [Page 172]). It must be highlighted that the butterflies in Dibang Valley are important not

only because they are essential pollinators, but also because they are key prey for other species, have cultural significance, and are vital for eco-tourism and have existence value.

SECTION 5: REVIEW OF AQUATIC BIODIVERSITY

Arunachal Pradesh's extensive river system sustains well over 250 species of fish from 105 genera, 34 families and 11 orders underscoring its evolutionary diversity (Bagra et al. 2009; Gurumayum et al. 2016). Of these, 32 species are endemic to the state (Gurumayum et al. 2016). Many fish species new to science have been reported from Arunachal Pradesh within a short span of time; most from the Siang, Noa-Dihing, Dibang and Subansiri rivers and their tributaries.

Underreporting fish species from Dibang Valley

In the Dibang River basin, Darshan et al. (2019) recorded 32 species, many of which were documented within the past 10 years, suggesting the potential of new discoveries from this species-rich region. However, the Report records only 12 species from the ZoI (including both Dri and Talõ basins) suggesting poor sampling. While the Report does plot the species accumulation curve for fish, the curve does not plateau indicating several undiscovered species in unsampled stream orders. Ichthyofaunal studies in Dibang Valley have been limited and only recently have researchers started to document the species for this region (see Appendix II for a list of 8 new species that have been described from this region within the last 10 years).

Dibang Valley harbours many endemic species, especially the highly sensitive, stenotopic Glyptoternoids like *Exostoma*, *Creteuchiloglanis*, *Parachiloglanis*, *Pseudolaguvia* and *Pseudocheineis*, but the Report only acknowledges one RET species (*Schizothorax richardsonii*). It further argues that other recorded species might not have global significance with reference to their threatened status, but they might have high regional importance. However, the fact is left out that the extent of the current knowledge of these species' ecology and conservation status is very limited making it difficult to make definitive assessments.

Threats to aquatic fauna underassessed

The Etalin HEP poses serious threats to the aquatic fauna of these high-altitude rivers which have not yet been adequately explored. The threats to migratory fish such as *Schizothorax* and *Tor* sp remain uncertain as there is inadequate knowledge on their migratory ranges. The location of the HEP at an altitude of 700 m is favourable for many "Intermediate Forms" of hill stream fishes including *Schizothorax*, some *Garra*, *Tor*, *Bangana*, *Neolissochilus*. These are potential "Cold Water Fishes" that migrate downstream to mid-to-low altitudes during winter. No mitigation plans are recommended to allow for the upstream to downstream movement of these fish species through species-specific fish ladders which should be based on a thorough study of species biology.

Fishing through traditional traps has also been an important source of dietary protein for the indigenous Idu Mishmi people, the availability of which may be affected by the disturbance related to the HEP. Further, the delineated ZoI does not take into account that areas upstream and downstream of the dams that will be severely altered due to storage of sediments. Once the sediments are trapped, the river downstream will turn sediment hungry, eroding the banks' riverine areas and removing spawning gravels used by fish (International Hydropower Association, 2019).

Inadequate mitigation plan for hydrology

Hydrological monitoring of the streams to assess impacts of road construction, habitat modification and management of aquatic biodiversity, has not been recommended. This is critical to measure low flows and water quality changes pre- and post- dam activities (Page 15). In addition:

- The mitigation plans recommend that critical minimum flows (e-flows) should be maintained in all the streams and the main river. However, they fail to detail the methodology to estimate the same (at different time of a year) for streams/ivers of different sizes and the fact that hydrological monitoring is crucial for deriving e-flows for different aquatic biota.
- There is a proposal to “*prevent the impact of road-cutting through the construction of culverts/small dams across all the streams cutting across by the proposed road is an effective mitigation plan*”. However this is only feasible for a few streams. Most first order streams do not get individual culverts and are diverted to the nearest culverts. This increases the runoff in the stream leading to higher erosion and bank instability (Page 200).
- The compound effects of ongoing landslides and excessive dumping of debris directly into the rivers due to highway construction that have already compromised stream health and productivity of other aquatic habitats, should be explicitly accounted for (Sassa and Canuti, 2009).
- The impact of the project on long-term sediment dynamics downstream are not addressed although short term effects of increased sedimentation during construction and immediate post-construction phase are addressed to some extent.
- The impact of altered flow and sediment regimes on riparian vegetation are not addressed.

SECTION 6: REVIEW OF AVIFAUNA

Even within Arunachal Pradesh (which has the second highest number of bird species globally), Dibang Valley is special for its birds. Four hundred and thirty-four species have been reported from Dibang Valley (see Appendix III for a checklist of birds of Dibang Valley). The Black-headed Greenfinch (*Chloris ambigua*) and Elliot’s Laughingthrush (*Trochalopteron elliotii*) are recent new records for India and the Rusty-throated Wren-Babbler (*Spelaornis badeigularis*) is a highly range-restricted and threatened species that was rediscovered from the area in 2004 after almost 60 years. The species is endemic to the region and found nowhere else in the world.

In addition, Arunachal Pradesh, including the Dibang Valley, has the highest richness of riverine birds on Earth. These species are entirely dependent on rivers and riverbank habitat of the Dibang and its tributaries. These discoveries and rediscoveries underscore that even relatively well-known faunal groups such as birds are poorly studied in Dibang Valley. While we found at least two identification errors in the photographs (Rosy Pipit has been wrongly identified as Olive-backed Pipit and Yellow-bellied Fairy Fantail, a common bird in this area, has been wrongly identified as the Yellow-throated Fulvetta), our major comments are as follows:

Missing the elevational migrants

The Report studied the birdlife along the Talõ and Dri rivers using point counts and line transects during February to May 2018. While the objective was to conduct a multi-season study, the actual field work

(February-May 2018) was done in a single season and is a relatively short time to assess birdlife of any Himalayan region, especially the species-rich Dibang Valley. First, most species in Arunachal Pradesh are elevational migrants, breeding at higher elevations and spending the winter lower down. Therefore, a study restricted to four months cannot provide a complete picture of the region's birdlife. Second, the Eastern Himalayan bird community is exceptionally diverse, and requires identification by both sight and sound. Over such a limited sampling period, it is likely that a significant proportion of species might not have been recorded by the study.

Underestimation of species, abundance and threatened species

The Report of 230 species from the study area is likely to be an underestimate, because of the lack of a multi-season study (as outlined above), and species present were not recorded during the survey because of poor detectability. The Report specifically outlines that "the vegetation was dense and the detectability (of birds) was less...". Given the low detection probability and large number of detections of singletons and doubletons, the Report did not incorporate a formal statistical technique that could potentially account for the number of species "missed". There are likely to be threatened species such as the Blyth's Tragopan and Beautiful Nuthatch in the area. Further surveys are required to systematically determine their presence in the landscape. No species accumulation curve given to show whether sampling was adequate.

On one hand the Report outlines that detectability was poor due to dense vegetation, yet goes ahead and classifies abundance of all birds that were recorded as very low or low. While the team has used point counts and line transects for the surveys, the number of repeats carried out have not been explicitly mentioned. The low abundances for birds inferred in the Report may be an artefact of low detection probability. Given low detection probability of species, abundance cannot be inferred without statistically accounting for the detection probability.

Contradictions and inadequate mitigation plans

The Report makes numerous statements that highlight high bird species richness and diversity of the region. One striking and key point is that even with this four-month study combined with effort/sampling limitations, the Report finds high bird species richness including endemics/range-restricted species within the ZoI and repeatedly stresses that it is critical/crucial to preserve these sites several times in The Report. Despite their own findings of the importance of biodiversity (in this case, birds), they suggest mitigation/conservation plans that clearly will not address the loss of these natural habitats and the direct impacts on bird populations. For instance, there are no mitigation measures suggested for the loss of habitat for riverine birds.

They also make contradictory assertions – where they admit that it is not possible to suggest any threatened and habitat specific conservation plans (which is true), and then subsequently suggest "*Habitat Rehabilitation and Restoration Plans*" that would purportedly enhance the overall habitat quality and benefit species of conservation significance.

Other limitations of habitat and species-specific plans

To mitigate or compensate the impacts on birds, the Report suggests the installation of nest boxes that the authors suggest will help the 32 cavity-nesting species of the 230 bird species recorded. This mitigation plan has several flaws, limitations, several of which are outlined below:

Cavity-nesting birds cannot just make do with some artificial nesting habitat, they need food, foraging areas and habitat to survive and breed. If the habitat (with nearly 2.8 lakh trees and other vegetation, associated features) are lost, birds will be directly impacted including the cavity-nesting species (which belong to a diversity of feeding guilds). The direct loss of the habitat/trees and submergence would reduce the population of birds. This cannot be compensated by only installing some nest boxes in adjacent areas.

The suggestion to augment only one type of breeding habitat (for cavity-nesting birds) by introducing nest boxes is an ecologically flawed idea as it may affect the population dynamics of other birds (assuming it is implemented and succeeds). There is no understanding of the abundances/population dynamics of birds in that area and no information on the existing availability of breeding habitat for cavity-nesting or other birds.

The 32 cavity-nesting species listed in the Report range from diurnal/nocturnal species, with diverse diet/foraging strategies and a range of body sizes. The birds range from tits, nuthatches, woodpeckers, barbets, to trogons and hornbills and raptors such as kestrels, hobby/owls. The design and placement of nest boxes for any particular species would require a much greater ecological understanding of the needs/requirements of specific species than one simple design based on differing entrance hole sizes catering to all species. The internal dimensions and specifications inside the nest box will also vary between different cavity-nesting species based on body size and other criteria.

The Report recommends that initially 400 such nest boxes should be set up in 2 locations (200 each) in forest patches around the staff colony and office premises. This proposal seems to be suggestive of an experimental framework and cannot be considered a mitigation strategy.

However, in other places, it is also suggested as a mitigatory measure with the idea that this would help cavity-nesting birds. This is misleading and incorrect. Till date, there is no successful project that shows the use and efficacy of nest boxes for hole-nesting bird species in India. In India, it has been tried for a handful of bird species in urban areas and in a few places for some hornbill species where years of ecological research have shown a limitation in the availability of natural cavities. Several of these projects especially for Asian forest hornbills have not yielded any significant results except in a couple of sites where it has needed careful and dedicated long-term engagement. In the case of hornbills or other large hole-nesting birds, installation and design of nest boxes is not an easy task, it requires multiple skills and equipment. In addition, nest boxes made of wood (as cursorily shown in the report) are also not at all ideal in places like Arunachal with high humidity and rainfall, where the boxes will rot after the first two years. Nest boxes often take a long time to be accepted and used by birds.

Many Environmental Management Plans of proposed hydel projects make this generic recommendation of installing nest boxes as a somewhat easy quick fix solution to compensate for the loss of birds/habitat. It would be very instructive to know of even a single example where this recommendation has been implemented in India in any project as a mitigatory measure.

SECTION 7: REVIEW OF MAMMALIAN BIODIVERSITY

Dibang Valley hosts an extremely rich mammalian assemblage with reports of as many as 75 species from the greater landscape of Dihang Dibang Biosphere Reserve (Choudhury, 2008). To assess the ZoI's mammalian diversity, the Report used a combination of camera traps and secondary sources. It claims evidence of 21 species within the ZoI from direct and indirect sources. Overall, flawed methodologies have been adopted to create an inadequate and incomplete assessment of the area's mammals and the HEP's impacts on them. In compiling data on mammals, the Report ignored key published checklists from the region (e.g. ZSI, 2006; Choudhury, 2008) instead opting for the EIA report (2015) that the FAC (2017) deemed "completely inadequate" leading to the commissioning of the current study (see Appendix IV for an complete checklist of mammals of Dibang Valley). They seem neither to have taken note of globally unique evolutionary phenomena reported from Dibang Valley (e.g. six different colour morphs of the Asiatic golden cat from the same region [Nijhawan et al., 2019]), nor the seven species of gliding squirrels, at least one of which (Mishmi Hill giant gliding squirrel) is endemic to the Dibang River basin (Krishna et al., 2016). The sections below discuss the various deficiencies and shortcomings in field and analytical methods, results and finally, the Report's conclusions on mitigation measures for mammals.

Flawed data collection methodologies

In Section 4.1.1.5 (Page 48), the Report states that "each camera was deployed for an average of 20-30 days" without explaining how this sampling period responds to the Objective (a) (Page 28) "covering multiple seasons". Firstly, a period of 20-30 days appears inadequate to sample a range of species, many of which are wide-ranging (Wearn and Glover-Kapfer, 2017). Secondly, there is no mention of how this sampling period is divided across different seasons as temporal replicates. Even if one assumes that 30 days were distributed equally across the two seasons identified in the report, a period of 15 days at any location is extremely unlikely to provide accurate estimates of species richness, distribution, and abundance, particularly for wide-ranging animals such as tiger, wild dog, and clouded leopard. Finally, mammals are known to change their distribution patterns based on resource availability in response to seasonal and altitudinal variation. Many mammals in Dibang Valley including Mishmi takin, red goral undertake altitudinal migrations (Choudhury 2008). Short survey durations restricted to specific seasons are sure to underestimate mammal presence.

The description provided under 'Random sampling' (Page 48) does not correspond to any acceptable ecological sampling method. No rigorous and robust inference can be drawn on species absence from a sampling approach that conducts 'opportunistic surveys based on information given by local people'. Relying entirely on the information provided by local people, though valuable, is likely to bias species capture probabilities, in particular for wide-ranging species such as large carnivores that may avoid human settlements (Schuette et al., 2013).

The Report claims to have conducted camera trap surveys in 53 sq. km of the 112 sq. km demarcated ZoI divided into 1x1 km grids (Page 48). This means that 48.18% of the ZoI was sampled. There are several concerning issues with the sampling methodology:

- All of the grids sampled are concentrated in the immediate vicinity of the river, human settlements and the district road. No grids higher up on mountain slopes within the ZoI were surveyed. This survey design is likely to have favoured the detection of species that are either found close to human settlements (e.g. rodents and small carnivores) or are more likely to visit

exposed riverbeds (such as otters and small cats), effectively excluding a range of species found farther away from human settlements.

- The Report does not provide the GPS coordinates of camera trap locations. However, Map 4.5 makes it clear that most of the cameras were placed within the roughly 32 of 112 grids, clustered close to the river, roads and settlements. The Report provides no rationale for purposefully selecting sampling grids and camera locations within them in areas known to be impacted by human presence, while leaving a significant majority of the ZoI unsurveyed. Furthermore, it is unclear how the Report concluded that an area of 53 sq. km was surveyed when so few grids have been effectively sampled.
- There is no mention of the minimum distance between two camera trap locations, therefore it is not possible to determine whether spatial autocorrelation in the photocapture data skewed estimates of the species richness and relative abundance.

A short survey duration coupled with non-representative sampling that unequally distributed sampling effort within a relatively small area (53 sq. km) render this study's findings on the study area's mammalian diversity inadequate. Yet, despite these serious methodological flaws, the Report **recorded 21 species of mammals, including the Critically Endangered Chinese Pangolin, and seven other RET species** pointing to the ZoI's importance for diverse, rare and threatened mammals.

Unscientific data analysis methods

The biased sampling strategy adopted in the Report does not meet the assumptions necessary to produce reliable estimates of species richness, distribution and abundance (Wearn and Glover-Kapfer, 2017). Consequently, richness and abundance analyses reported here (Page 122-132) carry little significance. Yet, if one assesses this section on its own merit, there are several concerns. Most importantly, the Report does not specify which method was employed to calculate species capture frequencies: 30-minute interval (O'Brien et al., 2003), 'independent encounter' (Rowcliffe et al., 2008) or another method. In addition to this key omission, there are following shortcomings:

- The Report does not consider differences in species detection probabilities and abundances (i.e., assigning the same weightage to captures of rare species with large home ranges, such as Asiatic wild dog and Asiatic black bear, and those with smaller home ranges, such as Himalayan palm civet and yellow-throated marten) when comparing species capture frequencies against one another. Meaningful comparisons in abundance across species cannot be made without accounting for inter-species variation (Sollman et al., 2013).
- No scientific reasoning is provided for setting the thresholds of abundance categories (very low to very high). No statistical tests are run either to test the significance of difference between the categories.
- Since the camera traps were only deployed for 20-30 days each (Page 48), it is unclear whether saturation (species accumulation) was reached in order to compile a comprehensive mammal checklist or to ascertain species richness, suggesting that the ZoI could contain many more species than were detected during this short and spatially restricted survey.

Flawed inference

The Report's claims of 'very low abundance' of mammals in the study area (Page 115) are unsubstantiated given the serious issues with data collection and analysis outlined above. Relative abundance index (RAI) is a poor estimator of abundance if species, survey and camera trap unit-species factors are not factored in (Sollman et al., 2013). The Report neither incorporates these factors nor

employs published methods such as the Random Encounter Model (Rowcliffe et al., 2008) or the Camera Trap Distance Sampling (Howe et al., 2017) used to produce reliable and robust estimates of species abundance from camera trapped data. An extensive long-term camera trap study in Dibang Valley that estimated species abundance using the Random Encounter Model (REM) showed that mammal abundances in Dibang's community forests were comparable to other tiger reserves in India with similar ecological carrying capacities (Nijhawan, 2018).

The stated rationale that since Dibang WLS is located 10 km from the ZoI (Page 116), the species found there should be excluded from the study area is at odds with widely known information on movement ecology and habitat preference of many of the area's species. Species such as Mishmi takin, red goral and Himalayan black bear are known to seasonally migrate between elevations, spending summers in higher altitudes and descending to lower elevation during winters (Choudhury 2008). Additionally, an aerial distance of 10 km is well within the limits of species with large home ranges especially big cats such as the tiger, Asiatic wild dog and clouded leopard that have been recorded in significant numbers in the region, even by WII's own scientists (Nijhawan, 2018; Adhikharimayum and Gopi, 2018).

Table 5.45 fails to mention the Chinese pangolin as Schedule I (Part I) species while Asiatic wild dog, smooth-coated otter and Himalayan black bear are included in Schedule II of the IWPA, meaning that these species are legally protected by provisions of the act. Additionally, Himalayan serow is classified as Near Threatened which the table also omits. Additionally, claims that threatened species such as spotted linsang and clouded leopard would not exist in the area (Page 117) are easily refutable as both have been reported from elevations lower than and similar to the ZoI in Dibang Valley (Nijhawan, 2018), Namdapha (Datta et al., 2008a; b) and Dampa Tiger Reserves (Singh and MacDonald, 2017).

Map 6.1 spatially displays the potential impacts of the proposed HEP divided into four categories from 'Very High' to 'Low'. This assessment compounds the serious methodological issues of data collection and analysis explained in prior sections. Thus, any results it generates are entirely unreliable. Beyond this, a striking pattern becomes clear if one compares Map 6.1 with Map 4.5. The grids identified as medium-to-high impact ('very high', 'high', 'medium') coincide with the grids where camera traps were placed. Therefore, if more extensive and longer camera trapping surveys had been conducted, species captures would have been higher. In sum, the impact assessment, and any mitigative measures by association, do not correspond to the true species abundance in the study area.

Mitigatory measures and impacts on mammals

The Report does not suggest any mitigatory measures for region's medium-to-large sized mammals impacted by the project, instead stating, "low abundance status of most of the species and few individuals of threatened species, along with predomination of forest and river habitat and absence of any critical habitat, it was not possible to suggest any threatened species and habitat specific conservation plan" (Page 185). The claim of absence of 'critical habitat' contradicts both the FAC's observations that "The type of forests appears to be predominantly Subtropical Evergreen broad-leaved forest and Subtropical rain... The vegetation is of multi-strata and can truly be said to be irreplaceable", and the fact Dibang Valley is part of a Global Biodiversity Hotspot – 36 such places that constitute 2.4% of the earth's surface but host 60% of all biodiversity on earth. The region is certainly critical habitat for several endangered and rare species (Choudhury 2008), which the Report fails to adequately document because of its flawed methodology.

The Report identifies only two areas within the region as being ‘ecologically sensitive’ – Dibang and Mehao WL sanctuaries – without providing any explanation of how only these were deemed ‘sensitive’ and the community forests were not. This directly contradicts a previous long-term study in Dibang Valley that showed that many community forests supported higher species richness and abundance for mammals than Dibang WLS (Nijhawan, 2018). Furthermore, and paradoxically, the Report concludes that “Nevertheless, continuous monitoring of movements of key mammalian fauna covering 10 km radius from the project study area is very important”, “[the] monitoring of tiger distribution and movements need to be continued in upper and lower Dibang Valley”, and “it is essential that long-term monitoring and conservation efforts are planned particularly for species of conservation significance (Table 5.46) such as Mishmi Takin (endemic species), Alpine Musk Deer, Red goral, Clouded Leopard, Snow Leopard, Spotted Linsang, *in and around the study area*” [emphasis added]. These statements imply two assumptions: (1) that the HEP’s impacts will be experienced within 10 km of its radius, and (2) the species of conservation exist within or close to the study area. If these are indeed true, then by not adequately surveying the area inside the small ZoI and within 10 km of the proposed site, the Report has failed to present a comprehensive and reliable assessment of the study area’s biodiversity and the HEP’s impacts on it. In the same vein, one is hard-pressed to understand why the Report ignores the outcome of another long-term WII study (Adhikarimayum and Gopi, 2018) that found camera trap evidence of tigers within 10 km radius of the HEP site, if indeed the “10 km radius from the project study area is very important”. Finally, these statements on the need for a monitoring study contradict the report’s mandate which was to assess biodiversity to determine the potential impacts of HEP on it) *before* the construction of the HEP and not after.

SECTION 8: REVIEW OF HERPETOFAUNA

Amphibians and reptiles are declining globally due to habitat fragmentation, climate change and diseases (Gibbons 2000). Amphibians, in particular, show high rates of endemism and niche specialisation (Hu et al., 2012; Brown et al., 2016). Both groups feed on insects, fish, small mammals and other reptiles, and provide key ecosystem functions by distributing nutrients across and within different trophic levels. Population declines in either of these groups could potentially lead to a collapse in these nutrient transfer networks (Odum, 1971). Furthermore, such declines may lead to consequent population increases in groups that may be harmful to human health (e.g. insects and rodent pests), or decreases in predatory mammal and bird species that depend on herpetofauna (Aguilar et al., 2013; Hocking and Babbitt, 2014). Simple counts of species richness or diversity estimates do not capture the importance of such vital trophic interactions.

Dibang Valley has many undiscovered species

At least 95 species of herpetofauna have been reported from the Dibang river basin, including 48 amphibian, 12 lizard, and 34 snake species. (Borah and Bordoloi, 2004; Athreya and Sheth, 2016; Roy et al., 2018; Ohler et al., 2018). Several species that are found in neighbouring Myanmar and China are also found here (e.g., *Rhacophorus translineatus* and *Trimeresurus medoensis*). Recent studies in the Western Ghats have used molecular approaches to show that many species previously thought to be widespread across these mountains’ complex topography may in fact be entirely distinct species (Garg et al. 2017). This is extremely likely to be the case with herpetofauna in Dibang Valley as it’s numerous mountain ridges and deep valleys can act as barriers to dispersal, resulting in speciation and endemism (Wollenberg et al., 2008; Che et al., 2010). Here, scientists have recently identified 23 amphibian genera and 36 reptile genera A comparison with herpetofaunal data from across Northeastern India, and West-

Central Arunachal in particular, has indicated that these recently reported species from Dibang Valley, many of which are yet to formally be described, could be new to science. These include, but are not limited to, species within the following genera: *Amolops*, *Bufo*, *Cyrtodactylus*, *Duttaphrynus*, *Ingerana*, *Kurixalus*, *Microhyla*, *Nasutixalus*, *Oreolalax*, *Philautus*, *Rhacophorus*, *Scutigera*, *Theloderma*, and *Xenophrys*.

Underreporting due to inappropriate methodology and under-sampling

The report's documented evidence of 14 amphibian and 31 reptile species is significantly lower than the total number of species (95) reported in all previous studies in Dibang Valley conducted in habitats and elevation gradients comparable to that of the ZoI (Borah and Bordoloi 2004; Athreya and Sheth 2016; Roy et al. 2018; Ohler et al. 2018). Shockingly, the Report fails to record a *Bufo* species (an Indo-Chinese toad), locally called 'Pahu', which is extremely abundant in Dibang Valley even during the season when the fieldwork was conducted. Furthermore, the Report includes the following species in its checklist: *Feihyala vittatus*, *Occidozyga borealis*, *Clinotarsus alticola*, *Polypedates teraiensis*. This is highly suspicious because not only are these species strict monsoon-breeders, which would make it very difficult to encounter them during the non-monsoon fieldwork, but also because none of the other multi-year extensive surveys mentioned above have been able to confirm their presence in Dibang Valley. These and many more discrepancies are a result of very limited sampling in the non-breeding season which has led to an overall underestimation of the study area's herpetofaunal diversity. These are discussed in detail below:

- The fieldwork was conducted between February and May/June, that is, before the monsoon season. This is the incorrect season for herpetofaunal surveys in the region as a large majority of the Dibang Valley's herpetofauna breed during the monsoon, and are consequently most active during this period, increasing the probability of detection (Ahmed et al., 2009; Sarkar and Ray, 2010). In direct comparison, Roy et al. (2018) reported 41 species of amphibians from field surveys conducted across all seasons, including monsoon, over three consecutive years covering 24 km of transects along streams, forests, grasslands and wetlands across the Dibang River Basin. Roy et al. (2018) encountered a higher number of species because they sampled different types of habitat spread over a larger elevation range (200-3500m). The Report does not refer to this ground-breaking, comprehensive and highly relevant piece of work.
- The study did not survey all habitats present in the ZoI including the various successive stages of evergreen and temperate broad-leaved forest, grassland, rivers and streams, wetlands, and ponds (Roy et al., 2018). Instead, the sampling was conducted almost entirely along roads (Page 44). The Report does not provide a rationale for the focus on roads which are widely known to create disturbance and barriers to movement, in addition to being non-ideal habitats for herpetofauna (Carr and Fahrig, 2001; Bennett, 2017; Marsh et al., 2017). Ongoing highway construction between Anini and Etalin has further increased the level of disturbance in these already disturbed areas. Road surveys only allow for the detection of few ground-dwelling species that access these areas. Furthermore, only 38 km of road length was surveyed. Because these roads are present only on the left banks of the Dri and Talõ rivers, the survey missed 70 of the 112 sampling grids, effectively accessing roads in mere 37.5% of the 112 km² ZoI.
- Streams were not sampled thoroughly. Amphibians are water-dependent and are found in abundance in streams and other water bodies. Additionally, different stream orders host different amphibian communities with headwater streams being keystone habitats for montane amphibians (Gillespie et al. 2004; Stoddard et al. 2004; Ficetola et al. 2011). The study does not provide any

information on how and what type of streams were selected for surveys. Furthermore, the sampling time window of 5-10 minutes allotted for streams is much too short and is certain to have missed many rare and difficult to observe species.

- Tree frogs are severely under-sampled based on comparisons with studies in Dibang Valley and other parts of Arunachal Pradesh (Pawar and Birand 2001; Tesia and Bordoloi 2013; Roy et al. 2018). The largest land-cover in the ZoI are different types of forests and even these have not been surveyed thoroughly, casting doubts over the representativeness of the survey's findings.
- A sizeable population of Keeled box turtle *Cuora mouhotii* (IUCN Endangered), locally called 'Ichimbo', was recorded from forest patches between 200-1000m elevation range forest patches downstream of the project site (Ahmed and Roy, 2016). Predictive habitat suitability modelling has indicated that there is a very high likelihood of this species being present around Etalin which has similar habitat type and elevation range (Ahmed and Roy, 2016). Due to large-scale habitat change, this little-studied, extremely rare turtle may be pushed towards local extinction.

Even with the above-mentioned biases in sampling methodology, incorrect sampling season and inadequate survey locations and distances, **the study found 14 species underscoring the richness of the habitat in the project area.** In sum, since amphibian richness and abundance were not adequately sampled across the ZoI, the Report has understated the true impacts of the HEP on this globally threatened faunal group. Most of these herpetofaunal species remain data deficient according to the IUCN's Redlist data underscoring the importance of comprehensive studies to ascertain their ecology and conservation status.

SECTION 9: LIMITED CRITIQUE OF SOCIO-CULTURAL SURVEYS

Overall, the parts of the Report related to assessing socio-cultural impacts of the HEP and corresponding mitigatory measures show a lack of understanding of complex socio-ecological dynamics and nature-culture interdependencies. The Report does not reference recent and widely-distributed published work on the relations between the Idu Mishmi, the natural environment and transboundary issues (e.g. Aiyadurai, 2016; Aiyadurai and Lee, 2017; Aiyadurai, 2018), ethnobiological knowledge (e.g. Ghosh et al., 2014) and publications by Idu Mishmi scholars on local belief systems (e.g. Mihi et al., 2018). While a detailed sociological critique of the Report is outside the scope of this review, the points below highlight some particularly concerning trends:

- **Basic population figures are incorrect:** The Report cites the 2011 All India Census incorrectly in stating that the Idu Mishmi population of Dibang Valley is 8004 (Page 34). While the 2011 Census does not specify the Idu Mishmi population of the district, it clearly mentions that 71.23% (5701) of the total population is Scheduled Tribe (ST). Since the Idu Mishmi is the primary ST in Dibang Valley, a large majority of the 5701 STs are likely to be Idu.
- **Flawed understanding of local livelihoods:** In various places, the Report claims that "about 38.2% of PAFs [Project affected families] are dependent on forests and their resources, as being their primary source of income....", and "only 4% of the people are dependent on agriculture for their livelihood income..." (Page 136; Table 5.55; Figure 5.10). These data and resulting conclusions seem to have a flawed idea of local livelihoods, assuming that those who are engaged in 'non-forest' livelihoods such as 'business', 'contract', 'labour' etc., do not simultaneously depend on forest-based products. Local livelihoods in Arunachal Pradesh cannot be categorized neatly into one form or the other as people depend upon a variety of sources throughout the year. Thus,

people who have been categorised as dependent upon 'contract', 'labour' etc, may also be dependent upon forest-based products as well as agriculture at different times of the year to supplement their food and cultural needs. Based on the report's own findings that 86.3% of those surveyed were involved in NTFP collection (Page 139), even the households characterised as being dependent on government jobs are sure to be involved in both agriculture and extraction of forest products at different times of the year. Finally, years of social science research has shown that livelihood strategies in traditional societies aren't simply a source of cash income, they carry complex socio-cultural meanings and purposes which the Report entirely ignores (Shackleton et al., 2011; Singh et al., 2017; Singh et al., 2018).

- **Outdated perspectives on *jhum* agriculture:** The Report asserts that “*jhum* agriculture or shifting cultivation is known for causing loss of forest cover and associated biodiversity values” (Page 136). This is an outdated and flawed statement that is not supported by research within the last 50 years. Many of these misconceptions around *jhum* have been dispelled by seminal work over the years (Conklin, 1954; Ramakrishnan, 1990, Mandal and Raman, 2016). Contrary to state policies targeting *jhum*, long-term research on medium to long-fallow *jhum* (10-15 fallow years) has found that these systems contribute substantially to subsistence livelihoods while being environmentally sustainable and supporting rich biodiversity within fallows. Recent work on *jhum* among the Adi tribe in the nearby Siang valley highlights the importance of *jhum* in providing direct and indirect benefits as well as being a critical resource for poorer families who may not have alternate sources of income (Teegalapalli, 2017; Datta-Roy, 2019). For communities practising shifting cultivation, it is not merely a system of cultivation, but a socio-cultural activity that provides meaning to land and reifies individual and group identities.
- **Undermining the local importance of mithun:** The Report notes “people prefer wild meat but tend to consume domestic meat more often” (Page 143) highlighting the importance of domestic meat in the lives of Idu Mishmi. However, it entirely fails to mention that one of the most important components of domestic meat (in terms of biomass as well as socio-cultural aspects) is the mithun (*Bos frontalis*). An adult mithun weighs about 500 kg and its meat forms a key source of protein for remote villages. The mithun holds a strong cultural meaning for the various native peoples of Northeast India and continues to be used as a token of currency and social status. They are sacrificed during specific festivals, such as the Idu festival of *reh*, making them an unalienable component of socio-cultural lives as well as an important part of local economy (Nijhawan, 2018). Mithuns are free-ranging, utilising the forests around the village with occasional visits to the village. The HEP, in particular the displacement of entire villages, will have significant impacts on the mithun. In response to local people's concerns about impacts on mithun ('loss of grazing land for the mithun' in Table 5.57), the Report proposes mitigation measures that seem to have no relevance for the free-ranging mithuns in the area (e.g. “All the three action plans will be implemented and developed within the village Gaucher land (land allotted for grazing)”. The proposal to plant nutritional grasses in additional parcels of land ignores published research that mithun are browsers that depend on 42-60 distinct species of naturally occurring trees, lianas, shrubs and herbs (Taba et al., 2015; Jamir and Khare, 2018). Does the Report envision re-creating these entire forest ecosystems for the mithun?
- **Undermining the impacts of migrant labour:** There appears to be a minimization of the negative impacts of the project and local concerns about them throughout the Report (e.g. various repetitions in the top half of Table 5.57). For example, the expected increase in the local population from the influx of an estimated 12,000 additional in-migrants during the construction phase is incorrectly reported to be 150% given that the entire population of the district is 8004. The many serious socio-cultural, economic and safety issues resulting from such enormous and sudden demographic changes are lumped under “Cultural Issues” (Table 7.22) leaving the

mitigation up to “high-level village committees”. In doing so, the Report seems to be arguing that issues of local sovereignty and women’s safety (Table 7.22) are apparently cultural matters. What exactly will the User Agency and the new “committees” do to address these very serious threats to local safety and well-being is left completely unanswered.

- **Contradictory mitigation proposals:** The Report recommends that villagers should be encouraged to do “large scale vegetable and fruit gardening” (Table 7.17). In principle, an attention to the importance of generating alternative sources of local income is to be appreciated. However, such recommendations directly contradict the mitigation measures the Report proposes for conservation of biodiversity in earlier sections such as compensatory afforestation (Page 183). This implies that biodiversity faces direct impacts not only from habitat loss due to the project but also due to “large-scale” agriculture. Furthermore, it refers to these recommendations as “a kind of people’s biodiversity conservation plan and will also improve their life quality of villagers”. However, how an exercise like the People’s Biodiversity Register (PBR) qualifies as a mitigatory measure against loss of habitat and associated socio-cultural-economic benefits is left entirely unexplained. Finally, this raises an important question: shouldn’t a PBR exercise that documents local knowledge on biodiversity be conducted before and not after the project is developed?
- **Romanticised notions of livelihood transitions:** The Report lists various proposed activities (job opportunities, creating supplementary income generating sources, health care, improved education) as part of their Corporate Social Responsibility (Page 202; Section 7.7.2). They state that since providing jobs to members of all affected families is not possible, they will be supporting various income generating programmes to reduce the local dependency on natural resources. Some of the jobs listed and described as 'decent' are welder, fitter, plumber, electrician etc. The nature of jobs proposed are largely alien to most highland farmers and rests on the assumption that people can make an effortless transition to new livelihoods immediately after the shock of resettlement. Studies on dam-induced displacement have found that it has negative impacts on employment rate, income level, income resource, and overall well-being of people (Nusser, 2003; Baran and Myschowoda, 2009; Richter et al., 2010; Zou, 2011; Huang et al., 2018). Sudden restrictions on access to natural resources can have negative consequences and natural resources and subsistence agriculture can act as safety nets during transition periods, particularly for the most vulnerable sections of the society (Kura et al., 2017).

CONCLUSION

At the outset, it is important to highlight that the FAC (2017) does an apt assessment of the irreversible direct and the indirect impacts of the HEP on the area’s biodiversity: “The land in which the project is proposed is in pristine forests with riverine growth that once cut cannot be replaced”, and “[the] proposed project falls under the richest bio-geographical province of the Himalayan zone and falls under one of the mega biodiversity hotspots of the world.” Study after study has demonstrated that this region and its biodiversity is important both from a regional and a global perspective.

The Report suffers from several technical shortcomings. The sampling was done in a much smaller area (the ZOI) than will see the direct and indirect impacts of the HEP. Even within the ZOI, not all grids were surveyed, ignoring the potentially disastrous impact of the HEP on yet undiscovered and endemic taxa. In the few sampling grids that were surveyed, unscientific and biased methods were often used

and not all elevations were sampled despite ample evidence that elevational gradients contribute to the high biodiversity in the Eastern Himalayas. Moreover, several groups of taxa were not surveyed, including numerous insect orders such as Diptera, Orthoptera, and arthropod species such as scorpions, and pseudoscorpions, and other taxa such as crustaceans, molluscs, and protozoans. Excluding highly diverse taxa such as insects and arthropods underestimates biodiversity values. Finally, traditional ecological knowledge and the intricacies of nature-human relations of the local people that inhabit and use these landscapes has been consistently undervalued. Importantly, the report seemed to have ignored a large majority of high quality relevant published literature on Dibang Valley's ecology, geology and anthropology. Overall, this has resulted in serious discrepancies in assessing the true biodiversity value of the impacted area, including the many RET/endemic species.

Incomplete and inaccurate data lead to an erroneous and inadequate assessment of the impact potential of the proposed HEP on biodiversity. The assessment of impact potential was not based on peer-reviewed methodologies and did not account for the grids that were not sampled, underestimated the biodiversity impacts of the proposed HEP. Moreover, the estimated potential impacts do not account for the loss of feedbacks between trophic levels due to the loss of certain keystone species or habitats. This can and will have far-reaching consequences for the overall stability and resilience of the ecosystem. Trophic interactions are the result of millions of years of evolutionary processes and do not stabilize in a short time when disrupted. Thus, the impact potential reported cannot be used to assess the appropriate mitigatory measures on the potential damage to wildlife and habitat.

The Report's claim of 38.2% of the PAFs being dependent upon forest resources is an underestimate and does not consider the entire range of services that are utilised by forest-dependent communities. Decades of social science research has shown that livelihood strategies in traditional societies are not simply means for providing cash income, but that they also carry complex socio-cultural meanings and values, which the Report entirely ignores.

Despite the many methodological flaws, and that the study was shortened to 4 months (for all taxa, and socio-cultural impact, despite the FAC mandate for a multi-season replicated study), the Report still makes it evident that Dibang Valley is exceptionally rich in biodiversity as every grid cell that was sampled, contained at least one RET/endemic species. It is then shocking that based on these results, and without a mandate, the Report goes on to prepare a Wildlife Conservation Plan taking the HEP as a given. All this without adequately evaluating or discussing the nature of impacts on specific groups, in particular, the Critically Endangered, Endangered, Vulnerable, range-restricted and endemics species that were recorded by the Report in the ZoI.

Data from several research studies show that the landscape is highly diverse. It is not difficult to see that a large fraction of this biodiversity will be impacted by any disturbance to the habitat. However, the recommendations of this report do not come close to mitigating the serious impacts of this HEP. The few mitigatory measures recommended for some specific faunal groups in the form of butterfly, reptile parks and nest boxes cannot be considered as well-designed ecologically meaningful measures.

The Report suggests these mitigation measures with the implicit assumption that they will work without any complications (e.g.: whether or not nest boxes will be accepted and accounting for durability given harshness of weather). It is regrettable that given the unique and extreme importance of the landscape to regional and global biodiversity, the study does not even attempt to outline relevant and viable mitigation measures and extent of damage to wildlife habitats, ecosystem services, and the local people. For instance, no mitigatory measures are provided for the loss of habitat for riverine birds, despite

Dibang Valley harbouring some of the highest riverine bird diversity in the world. In other cases, the suggested mitigatory measures assume a patronizing attitude towards the local Idu Mishmi people and emerge from a lack of a nuanced understanding of socio-cultural dynamics and interdependencies between people and the natural environment. The Report recommends large-scale agriculture and a shift to cash crops, without considering the impact of such large scale (possibly monoculture) cultivation on biodiversity, local livelihoods and well-being. In the same vein, the conclusions on the impacts on mithun are short-sighted, and underestimate the cultural significance of these animals. By not accounting for a large influx of labourers and their impacts on the landscape, the wildlife habitat, and on the cultural identity, health and wellbeing of the local people, the Report undermines the impact of this project on multiple fronts.

Finally, and crucially, studies that inform high-level decision-making on historically significant projects, such as the Etalin HEP which would be the largest hydropower project in the country, must go through a transparent and scientifically recognised peer-reviewed process given the pitfalls, numerous discrepancies and gaps highlighted in this review. Such decisions have irreversible impacts on lives, livelihoods and the environment.

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APPENDIX I: CHECKLIST OF BUTTERFLIES OF DIBANG VALLEY (Gogoi 2020 unpublished data)

| Sl no. | Family | Common name | Scientific name |
|---------------|---------------|-------------------------|---|
| 1 | Papilionidae | Common Rose | <i>Pachliopta aristolochiae aristolochiae</i> |
| 2 | Papilionidae | Common Birdwing | <i>Troides helena cerberus</i> |
| 3 | Papilionidae | Golden Birdwing | <i>Troides aeacus aeacus</i> |
| 4 | Papilionidae | Common Batwing | <i>Atrophaneura varuna astorion</i> |
| 5 | Papilionidae | Lesser Batwing | <i>Atrophaneura aidoneus</i> |
| 6 | Papilionidae | De Nicéville's Windmill | <i>Byasa polla</i> |
| 7 | Papilionidae | Common Windmill | <i>Byasa polyeuctes polyeuctes</i> |
| 8 | Papilionidae | Great Windmill | <i>Byasa dasarada dasarada</i> |
| 9 | Papilionidae | Tawny Mime | <i>Papilio agestor agestor</i> |
| 10 | Papilionidae | Lesser Mime | <i>Papilio epycides epycides</i> |
| 11 | Papilionidae | Common Mime | <i>Papilio clytia clytia</i> |
| 12 | Papilionidae | Common Mormon | <i>Papilio polytes romulus</i> |
| 13 | Papilionidae | Lime Butterfly | <i>Papilio demoleus demoleus</i> |
| 14 | Papilionidae | Common Raven | <i>Papilio castor castor</i> |
| 15 | Papilionidae | Red Helen | <i>Papilio helenus helenus</i> |
| 16 | Papilionidae | Yellow Helen | <i>Papilio nephelus chaon</i> |
| 17 | Papilionidae | Great Mormon | <i>Papilio memnon agenor</i> |
| 18 | Papilionidae | Spangle | <i>Papilio protenor euprotenor</i> |
| 19 | Papilionidae | Redbreast | <i>Papilio alcmenor alcmenor</i> |
| 20 | Papilionidae | Common Peacock | <i>Papilio polyctor ganesa</i> |
| 21 | Papilionidae | Paris Peacock | <i>Papilio paris paris</i> |
| 22 | Papilionidae | Krishna Peacock | <i>Papilio krishna</i> |
| 23 | Papilionidae | Blue Peacock | <i>Papilio arcturus</i> |
| 24 | Papilionidae | Fivebar Swordtail | <i>Graphium antiphates pompilius</i> |
| 25 | Papilionidae | Fourbar Swordtail | <i>Graphium agetes agetes</i> |
| 26 | Papilionidae | Common Jay | <i>Graphium doson axion</i> |
| 27 | Papilionidae | Lesser Jay | <i>Graphium evemon albociliatis</i> |
| 28 | Papilionidae | Veined Jay | <i>Graphium chironides chironides</i> |
| 29 | Papilionidae | Tailed Jay | <i>Graphium agamemnon agamemnon</i> |
| 30 | Papilionidae | Common Bluebottle | <i>Graphium sarpedon sarpedon</i> |
| 31 | Papilionidae | Great Zebra | <i>Graphium xenocles xenocles</i> |
| 32 | Papilionidae | White Dragontail | <i>Lamproptera curius curius</i> |
| 33 | Papilionidae | Green Dragontail | <i>Lamproptera meges indistincta</i> |
| 34 | Papilionidae | Brown Gorgon | <i>Meandrusa lachinus lachinus</i> |
| 35 | Papilionidae | Yellow Gorgon | <i>Meandrusa payeni evan</i> |
| 36 | Pieridae | One-Spot Grass Yellow | <i>Eurema andersoni andersoni</i> |
| 37 | Pieridae | Three-Spot Grass Yellow | <i>Eurema blanda silhetana</i> |
| 38 | Pieridae | Small Grass Yellow | <i>Eurema brigitta rubella</i> |
| 39 | Pieridae | Common Grass Yellow | <i>Eurema hecabe hecabe</i> |
| 40 | Pieridae | Tree Yellow | <i>Gandaca harina assamica</i> |
| 41 | Pieridae | Tibetan Brimstone | <i>Gonepteryx amintha thibetana</i> |
| 42 | Pieridae | Tailed Sulphur | <i>Dercas verhuelli doubledayi</i> |

| | | | |
|----|------------|-----------------------------|---------------------------------------|
| 43 | Pieridae | Common Emigrant | <i>Catopsilia pomona pomona</i> |
| 44 | Pieridae | Mottled Emigrant | <i>Catopsilia pyranthe pyranthe</i> |
| 45 | Pieridae | Dark Clouded Yellow | <i>Colias fieldii fieldii</i> |
| 46 | Pieridae | Yellow Orange Tip | <i>Ixias pyrene familiaris</i> |
| 47 | Pieridae | Pale Wanderer | <i>Pareronia avatar avatar</i> |
| 48 | Pieridae | Chocolate Albatross | <i>Appias lyncida hippoides</i> |
| 49 | Pieridae | Orange Albatross | <i>Appias nero galba</i> |
| 50 | Pieridae | Common Albatross | <i>Appias albina darada</i> |
| 51 | Pieridae | Spot Puffin | <i>Appias lalage lalage</i> |
| 52 | Pieridae | Plain Puffin | <i>Appias indra indra</i> |
| 53 | Pieridae | Large Cabbage White | <i>Pieris brassicae</i> |
| 54 | Pieridae | Bhutan Blackvein | <i>Aporia harrietae</i> |
| 55 | Pieridae | Green-veined White | <i>Pieris napi montana</i> |
| 56 | Pieridae | Indian Cabbage White | <i>Pieris canidia indica</i> |
| 57 | Pieridae | Lesser Gull | <i>Cepora nadina nadina</i> |
| 58 | Pieridae | Common Gull | <i>Cepora nerissa</i> |
| 59 | Pieridae | Spotted Sawtooth | <i>Prioneris thestylis thestylis</i> |
| 60 | Pieridae | Redspot Sawtooth | <i>Prioneris clemathe</i> |
| 61 | Pieridae | Hill Jezebel | <i>Delias belladonna lugens</i> |
| 62 | Pieridae | Dark Jezebel | <i>Delias berinda</i> |
| 63 | Pieridae | Pale Jezebel | <i>Delias sanaca</i> |
| 64 | Pieridae | Red-breast Jezebel | <i>Delias acalis pyramus</i> |
| 65 | Pieridae | Yellow Jezebel | <i>Delias agostina agostina</i> |
| 66 | Lycaenidae | Angled Sunbeam | <i>Curetis dentata dentata</i> |
| 67 | Lycaenidae | Forest Pierrot | <i>Taraka hamada mendesia</i> |
| 68 | Lycaenidae | Straight Pierrot | <i>Caleta roxus roxana</i> |
| 69 | Lycaenidae | Elbowed Pierrot | <i>Caleta elna noliteia</i> |
| 70 | Lycaenidae | Common Pierrot | <i>Castalius rosimon rosimon</i> |
| 71 | Lycaenidae | Balkan Pierrot | <i>Tarucus balkanicus</i> |
| 72 | Lycaenidae | Silver Royal | <i>Ancema blanka</i> |
| 73 | Lycaenidae | Bi-Spot Royal | <i>Ancema ctesia ctesia</i> |
| 74 | Lycaenidae | Chocolate Royal | <i>Remelana jangala ravata</i> |
| 75 | Lycaenidae | Centaur Oakblue | <i>Arhopala centaurus pirithous</i> |
| 76 | Lycaenidae | Hooked Oakblue | <i>Arhopala paramuta</i> |
| 77 | Lycaenidae | Variogated Plushblue | <i>Flos adriana</i> |
| 78 | Lycaenidae | Spangled Plushblue | <i>Flos asoka</i> |
| 79 | Lycaenidae | Common Acacia Blue | <i>Surendra vivarna</i> |
| 80 | Lycaenidae | Silver Streaked Acacia Blue | <i>Zinaspa todara distorta</i> |
| 81 | Lycaenidae | Yamfly | <i>Loxura atymnus continentalis</i> |
| 82 | Lycaenidae | Branded Yamfly | <i>Yasoda tripunctata tripunctata</i> |
| 83 | Lycaenidae | Common Imperial | <i>Cheritra freja freja</i> |
| 84 | Lycaenidae | Blue Imperial | <i>Ticherra acte</i> |
| 85 | Lycaenidae | Common Tit | <i>Hypolycaena erylus himavantus</i> |
| 86 | Lycaenidae | Blue Tit | <i>Chliaria kina cachara</i> |
| 87 | Lycaenidae | Fluffy Tit | <i>Zeltus amasa</i> |
| 88 | Lycaenidae | Common Flash | <i>Rapala nissa ratna</i> |

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| 89 | Lycaenidae | Slate Flash | <i>Rapala manea schistacea</i> |
| 90 | - | Copper Flash | <i>Rapala pheritima</i> |
| 91 | Lycaenidae | - | <i>Eusaspa pavo</i> |
| 92 | Lycaenidae | - | <i>Ahlbergia sp</i> |
| 93 | Lycaenidae | Common Tinsel | <i>Catapocilma elegans</i> |
| 94 | Lycaenidae | Evans Silverline | <i>Spindasis evansii</i> |
| 95 | Lycaenidae | Khaki Silverline | <i>Spindasis rukmini</i> |
| 96 | Lycaenidae | Long-banded Silverline | <i>Spindasis lohita himalayanus</i> |
| 97 | Lycaenidae | Chinese Silverline | <i>Spindasis zhengweilie</i> |
| 98 | Lycaenidae | Purple Sapphire | <i>Heliophorus epicles</i> |
| 99 | Lycaenidae | Green Sapphire | <i>Heliophorus moorei</i> |
| 100 | Lycaenidae | Golden Sapphire | <i>Heliophorus brahma major</i> |
| 101 | Lycaenidae | Common Ciliate Blue | <i>Anthene emolus emolus</i> |
| 102 | Lycaenidae | Pointed Cilate Blue | <i>Anthene lycaenina lycaenina</i> |
| 103 | Lycaenidae | Zebra Blue | <i>Leptotes plinius plinius</i> |
| 104 | Lycaenidae | Large-4-Lineblue | <i>Nacaduba pactolus continentalis</i> |
| 105 | Lycaenidae | Pale-4-Lineblue | <i>Nacaduba hermus nabo</i> |
| 106 | Lycaenidae | Transparent 6-Lineblue | <i>Nacaduba kurava euplea</i> |
| 107 | Lycaenidae | Banded Lineblue | <i>Prosotas aluta coelestis</i> |
| 108 | Lycaenidae | Common Lineblue | <i>Prosotas nora nora</i> |
| 109 | Lycaenidae | Pointed Lineblue | <i>Lonolyce helicon merguiana</i> |
| 110 | Lycaenidae | Dingy Lineblue | <i>Petrelaea dana</i> |
| 111 | Lycaenidae | Common Cerulean | <i>Jamides celeno celeno</i> |
| 112 | Lycaenidae | Dark Cerulean | <i>Jamides bochus bochus</i> |
| 113 | Lycaenidae | Glistening Cerulean | <i>Jamides elpis pseudelpis</i> |
| 114 | Lycaenidae | Metallic Cerulean | <i>Jamides alecto eurysaces</i> |
| 115 | Lycaenidae | Forget-me-not | <i>Catochrysops strabo srrabo</i> |
| 116 | Lycaenidae | Peablue | <i>Lampides boeticus</i> |
| 117 | Lycaenidae | Pale Spark | <i>Sinthusia virga</i> |
| 118 | Lycaenidae | Dark Grass Blue | <i>Zizeeria karsandra</i> |
| 119 | Lycaenidae | Pale Grass Blue | <i>Pseudozizeeria maha maha</i> |
| 120 | Lycaenidae | Eastern Grass Jewel | <i>Freyeria putli</i> |
| 121 | Lycaenidae | Lesser Grass Blue | <i>Zizina otis otis</i> |
| 122 | Lycaenidae | Malayan | <i>Megisba malaya</i> |
| 123 | Lycaenidae | Common Hedge Blue | <i>Acytolepis puspa gisca</i> |
| 124 | Lycaenidae | Pale Hedge Blue | <i>Udara cardia dilecta</i> |
| 125 | Lycaenidae | Albocerulean | <i>Udara albocaerulea</i> |
| 126 | Lycaenidae | Plain Hedge Blue | <i>Celastrina lavendularis limbata</i> |
| 127 | Lycaenidae | Hill Hedge Blue | <i>Celastrina argiolus sikkima</i> |
| 128 | Lycaenidae | White banded Hedgeblue | <i>Lycaenopsis transpectus</i> |
| 129 | Lycaenidae | Marginated Hedgeblue | <i>Celatoxia marginata</i> |
| 130 | Lycaenidae | Large Hedge Blue | <i>Celastrina huegelii oreana</i> |
| 131 | Lycaenidae | Moore's Cupid | <i>Shijimia moorei</i> |
| 132 | Lycaenidae | False Tibetan Cupid | <i>Tongeia pseudozuthus</i> |
| 133 | Lycaenidae | Lime Blue | <i>Chilades laius laius</i> |
| 134 | Lycaenidae | Blue Posy | <i>Drupadia scaeva</i> |

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| 135 | Riodinidae | Dark Judy | <i>Abisara fylla</i> |
| 136 | Riodinidae | Punchinello | <i>Zemeros flegyas indicus</i> |
| 137 | Riodinidae | Mixed Punch | <i>Doona ouida</i> |
| 138 | Riodinidae | Lesser Punch | <i>Dodona dipoea</i> |
| 139 | Riodinidae | Tailed Punch | <i>Dodona eugens</i> |
| 140 | Riodinidae | Striped Punch | <i>Dodonia adonira naga</i> |
| 141 | Nymphalidae | Club Beak | <i>Libythea myrrha sanguinalis</i> |
| 142 | Nymphalidae | Common Beak | <i>Libythea lepita lepita</i> |
| 143 | Nymphalidae | Striped Tiger | <i>Danaus genutia</i> |
| 144 | Nymphalidae | Blue Tiger | <i>Tirumala limniace mutina</i> |
| 145 | Nymphalidae | Dark Blue Tiger | <i>Tirumala septentrionis</i> |
| 146 | Nymphalidae | Glassy Tiger | <i>Parantica aglea melanoides</i> |
| 147 | Nymphalidae | Chestnut Tiger | <i>Parantica sita</i> |
| 148 | Nymphalidae | Chocolate Tiger | <i>Parantica melaneus plateniston</i> |
| 149 | Nymphalidae | Striped Blue Crow | <i>Euploea mulciber mulciber</i> |
| 150 | Nymphalidae | Long-branded Blue Crow | <i>Euploea algea deione</i> |
| 151 | Nymphalidae | Magpie Crow | <i>Euploea radamanthus radamanthus</i> |
| 152 | Nymphalidae | Common Nawab | <i>Polyura athamas athamas</i> |
| 153 | Nymphalidae | Great Nawab | <i>Polyura eudamippus eudamippus</i> |
| 154 | Nymphalidae | Pallid Nawab | <i>Polyura arja arja</i> |
| 155 | Nymphalidae | Stately Nawab | <i>Polyura dolon</i> |
| 156 | Nymphalidae | Tawny Rajah | <i>Charaxes bernardus hierax</i> |
| 157 | Nymphalidae | Scarce Tawny Rajah | <i>Charaxes aristogiton</i> |
| 158 | Nymphalidae | Variiegated Rajah | <i>Charaxes kahruba</i> |
| 159 | Nymphalidae | Yellow Rajah | <i>Charaxes marmax marmax</i> |
| 160 | Nymphalidae | Jungle Glory | <i>Thaumantis diores diores</i> |
| 161 | Nymphalidae | Common Faun | <i>Faunis canens</i> |
| 162 | Nymphalidae | Common Duffer | <i>Discophora sondaica</i> |
| 163 | Nymphalidae | Common Evening Brown | <i>Melanitis leda</i> |
| 164 | Nymphalidae | Bamboo Treebrown | <i>Lethe europa niladana</i> |
| 165 | Nymphalidae | Common Red Forester | <i>Lethe mekara zuchara</i> |
| 166 | Nymphalidae | Angled Red Forester | <i>Lethe chandica flanona</i> |
| 167 | Nymphalidae | Tailed Red Forester | <i>Lethe sinorix</i> |
| 168 | Nymphalidae | Blue Forester | <i>Lethe scanda</i> |
| 169 | Nymphalidae | Common Forester | <i>Lethe insana</i> |
| 170 | Nymphalidae | Dull Forester | <i>Lethe gulnihal</i> |
| 171 | Nymphalidae | Brown Forester | <i>Lethe serbonis</i> |
| 172 | Nymphalidae | Common Woodbrown | <i>Lethe sidonis sidonis</i> |
| 173 | Nymphalidae | Barred Woodbrown | <i>Lethe maitrya</i> |
| 174 | Nymphalidae | Yellow Woodbrown | <i>Lethe nicetas</i> |
| 175 | Nymphalidae | Moeller's Silverfork | <i>Lethe moelleri</i> |
| 176 | Nymphalidae | Small Goldenfork | <i>Lethe atkinsonia</i> |
| 177 | Nymphalidae | Large Goldenfork | <i>Lethe goalpara</i> |
| 178 | Nymphalidae | Single Silverstripe | <i>Lethe ramadeva</i> |
| 179 | Nymphalidae | Lilacfork | <i>Lethe sura</i> |
| 180 | Nymphalidae | Scarce Labyrinth | <i>Neope pulahina</i> |

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| 181 | Nymphalidae | Dusky Labyrinth | <i>Neope yama</i> |
| 182 | Nymphalidae | Veined Labyrinth | <i>Neope pulaha</i> |
| 183 | Nymphalidae | Chumbi Wall | <i>Chonala masoni</i> |
| 184 | Nymphalidae | Large Tawny Wall | <i>Rhapticera satricus</i> |
| 185 | Nymphalidae | Dusky Diadem | <i>Ethope himachala</i> |
| 186 | Nymphalidae | Yellow Owl | <i>Neorina hilda</i> |
| 187 | Nymphalidae | Yellow Kaiser | <i>Penthema lisarda lisarda</i> |
| 188 | Nymphalidae | Common Palmfly | <i>Elymnias hypermenestra undularis</i> |
| 189 | Nymphalidae | Spotted Palmfly | <i>Elymnias malelas malelas</i> |
| 190 | Nymphalidae | Whitebar Bushbrown | <i>Mycalesis anaxias</i> |
| 191 | Nymphalidae | Watson's Bushbrown | <i>Mycalesis adamsoni</i> |
| 192 | Nymphalidae | Plain Busbrown | <i>Mycalesis malsarida</i> |
| 193 | Nymphalidae | Common Bushbrown | <i>Mycalesis perseus blasius</i> |
| 194 | Nymphalidae | Dark-brand Bushbrown | <i>Mycalesis mineus mineus</i> |
| 195 | Nymphalidae | Long-brand Bushbrown | <i>Mycalesis visala visala</i> |
| 196 | Nymphalidae | Salmon-branded Bushbrown | <i>Mycalesis misenus misenus</i> |
| 197 | Nymphalidae | Bright-eye Bushbrown | <i>Mycalesis nicotia</i> |
| 198 | Nymphalidae | Nigger | <i>Orsotrioena medus medus</i> |
| 199 | Nymphalidae | Striped Ringlet | <i>Ragadia crisilda crisilda</i> |
| 200 | Nymphalidae | Dark Catseye | <i>Zipoetis scylax</i> |
| 201 | Nymphalidae | Mottled Argus | <i>Hemadara narasingha</i> |
| 202 | Nymphalidae | Himalayan Fivering | <i>Ypthima parasakra</i> |
| 203 | Nymphalidae | Large Threering | <i>Ypthima newara?</i> |
| 204 | Nymphalidae | Common Fiverring | <i>Ypthima baldus baldus</i> |
| 205 | Nymphalidae | Ring sp. | <i>Ypthima sp.</i> |
| 206 | Nymphalidae | Roy's Argus | <i>Callerebia dibangensis</i> |
| 207 | Nymphalidae | Pallid Argus | <i>Callerebia scanda</i> |
| 208 | Nymphalidae | Argus sp | <i>Callerebia sp.</i> |
| 209 | Nymphalidae | Doherty's Satyr | <i>Aulocera loha</i> |
| 210 | Nymphalidae | Yellow Coster | <i>Acraea issoria issoria</i> |
| 211 | Nymphalidae | Tawny Coster | <i>Acraea violae</i> |
| 212 | Nymphalidae | Red Lacewing | <i>Cethosia biblis tisamena</i> |
| 213 | Nymphalidae | Leopard Lacewing | <i>Cethosia cyane cyane</i> |
| 214 | Nymphalidae | Indian Fritillary | <i>Argynnis hyperbius hyperbius</i> |
| 215 | Nymphalidae | Yellow Dryad | <i>Aemona amathusia</i> |
| 216 | Nymphalidae | Cruiser | <i>Vindula erota erota</i> |
| 217 | Nymphalidae | Common Yeoman | <i>Cirrochroa tyche mithila</i> |
| 218 | Nymphalidae | Large Yeoman | <i>Cirrochroa aoris aoris</i> |
| 219 | Nymphalidae | Rustic | <i>Cupha erymanthis lotis</i> |
| 220 | Nymphalidae | Vagrant | <i>Vagrans egista sinha</i> |
| 221 | Nymphalidae | Common Leopard | <i>Phalanta phalantha phalantha</i> |
| 222 | Nymphalidae | Green Commodore | <i>Sumalia daraxa daraxa</i> |
| 223 | Nymphalidae | Commodore | <i>Auzakia danava danava</i> |
| 224 | Nymphalidae | White Commodore | <i>Parasarpa dudu dudu</i> |
| 225 | Nymphalidae | Scarce White Commodore | <i>Limenitis zulema</i> |
| 226 | Nymphalidae | Grey Commodore | <i>Bhagadatta austenia</i> |

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| 227 | Nymphalidae | Bicolour Commodore | <i>Parasarpa zayla</i> |
| 228 | Nymphalidae | Commander | <i>Moduza procris procris</i> |
| 229 | Nymphalidae | Studded Sergeant | <i>Athyma asura asura</i> |
| 230 | Nymphalidae | Himalayan Sergeant | <i>Athyma opalina orientalis</i> |
| 231 | Nymphalidae | Blackvein Sergeant | <i>Athyma ranga ranga</i> |
| 232 | Nymphalidae | Staff Sergeant | <i>Athyma selenophora selenophora</i> |
| 233 | Nymphalidae | Small Staff Sergeant | <i>Athyma zeroa zeroa</i> |
| 234 | Nymphalidae | Orange Staff Sergeant | <i>Athyma cama</i> |
| 235 | Nymphalidae | Common Lascar | <i>Pantoporia hordonia hordonia</i> |
| 236 | Nymphalidae | Perak Lascar | <i>Pantoporia perak</i> |
| 237 | Nymphalidae | Great Yellow Sailer | <i>Neptis radha radha</i> |
| 238 | Nymphalidae | Yellow Sailer | <i>Neptis ananta ochracea</i> |
| 239 | Nymphalidae | Small Yellow Sailer | <i>Neptis miah miah</i> |
| 240 | Nymphalidae | Pale Hockeystick Sailer | <i>Neptis manasa manasa</i> |
| 241 | Nymphalidae | Common Sailer | <i>Neptis hylas astola</i> |
| 242 | Nymphalidae | Creamy Sailer | <i>Neptis soma soma</i> |
| 243 | Nymphalidae | Sullied Sailer | <i>Neptis clinia susruta</i> |
| 244 | Nymphalidae | Pallas Sailer | <i>Neptis sappho</i> |
| 245 | Nymphalidae | Broad-banded Sailer | <i>Neptis sankara amba</i> |
| 246 | Nymphalidae | Dingy Sailer | <i>Neptis pseudovikasi</i> |
| 247 | Nymphalidae | Plain Sailer | <i>Neptis cartica cartica</i> |
| 248 | Nymphalidae | Pale Green Sailer | <i>Neptis zaida</i> |
| 249 | Nymphalidae | Short-banded Sailer | <i>Phaedyma columella ophiana</i> |
| 250 | Nymphalidae | Common Baron | <i>Euthalia aconthea</i> |
| 251 | Nymphalidae | Blue Baron | <i>Euthalia telchinia</i> |
| 252 | Nymphalidae | Gaudy Baron | <i>Euthalia lubentina</i> |
| 253 | Nymphalidae | French Duke | <i>Euthalia francaie</i> |
| 254 | Nymphalidae | Grand Duchess | <i>Euthalia patala</i> |
| 255 | Nymphalidae | White edge Blue Baron | <i>Euthalia phemius</i> |
| 256 | Nymphalidae | Dark Archduke | <i>Lexias dirtea khasiana</i> |
| 257 | Nymphalidae | Bronze Duke | <i>Euthalia nara</i> |
| 258 | Nymphalidae | - | <i>Limenitis rileyi</i> |
| 259 | Nymphalidae | Common Map | <i>Cyrestis thyodamas thyodamas</i> |
| 260 | Nymphalidae | Common Maplet | <i>Chersonesia risa</i> |
| 261 | Nymphalidae | Tabby | <i>Pseudergolis wedah</i> |
| 262 | Nymphalidae | Constable | <i>Dichorrhagia nesimachus</i> |
| 263 | Nymphalidae | Popinjay | <i>Stibochiona nicea</i> |
| 264 | Nymphalidae | Angled Castor | <i>Ariadne ariadne pallidior</i> |
| 265 | Nymphalidae | Common Castor | <i>Ariadne merione tapestrina</i> |
| 266 | Nymphalidae | Sergeant Emperor | <i>Mimathyma chevana</i> |
| 267 | Nymphalidae | Indian Purple Emperor | <i>Mimathyma ambica</i> |
| 268 | Nymphalidae | Courtesan | <i>Euripus nyctelius</i> |
| 269 | Nymphalidae | Circe | <i>Hestinalis nama</i> |
| 270 | Nymphalidae | Eastern Courtier | <i>Sephisa chandra</i> |
| 271 | Nymphalidae | Common Jester | <i>Symbrenthia lilaea khasiana</i> |
| 272 | Nymphalidae | Spotted Jester | <i>Symbrenthia hypselis cotanda</i> |

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| 273 | Nymphalidae | Indian Red Admiral | <i>Vanessa indica indica</i> |
| 274 | Nymphalidae | Painted Lady | <i>Vanessa cardui</i> |
| 275 | Nymphalidae | Blue Admiral | <i>Kaniska canace canace</i> |
| 276 | Nymphalidae | Black Prince | <i>Rohana parisatis</i> |
| 277 | Nymphalidae | Brown Prince | <i>Rohana parvata</i> |
| 278 | Nymphalidae | Chocolate Pansy | <i>Junonia iphita iphita</i> |
| 279 | Nymphalidae | Grey Pansy | <i>Junonia atlites</i> |
| 280 | Nymphalidae | Peacock Pansy | <i>Junonia almana almana</i> |
| 281 | Nymphalidae | Lemon Pansy | <i>Junonia lemonias lemonias</i> |
| 282 | Nymphalidae | Great Eggfly | <i>Hypolimnas bolina</i> |
| 283 | Nymphalidae | Orange Oakleaf | <i>Kallima inachus inachus</i> |
| 284 | Nymphalidae | Autumn Leaf | <i>Doleschallia bisaltide indica</i> |
| 285 | Nymphalidae | Panther | <i>Neurosigma doubledayi</i> |
| 286 | Nymphalidae | Abor Freak | <i>Callinaga aborica</i> |
| 287 | Nymphalidae | Tiger Brown | <i>Orinoma damaris</i> |
| 288 | Hesperiidae | Branded Orange Awlet | <i>Burara oedipodea aegina</i> |
| 289 | Hesperiidae | Orange Awlet | <i>Burara jaina vasundhara</i> |
| 290 | Hesperiidae | Small Green Awlet | <i>Burara amara</i> |
| 291 | Hesperiidae | Green Awlet | <i>Burara vasutana</i> |
| 292 | Hesperiidae | Pale Green Awlet | <i>Burara gomata gomata</i> |
| 293 | Hesperiidae | Slate Awl | <i>Hasora anura danda</i> |
| 294 | Hesperiidae | Common Awl | <i>Hasora badra badra</i> |
| 295 | Hesperiidae | Plain Banded Awl | <i>Hasora vita indica</i> |
| 296 | Hesperiidae | Common Banded Awl | <i>Hasora chromus</i> |
| 297 | Hesperiidae | White-banded Awl | <i>Hasora taminatus bhavara</i> |
| 298 | Hesperiidae | Brown Awl | <i>Badamia exclamationis</i> |
| 299 | Hesperiidae | Orange-tail Awl | <i>Bibasis sena sena</i> |
| 300 | Hesperiidae | Indian Awlking | <i>Choaspes benjaminii</i> |
| 301 | Hesperiidae | Common Spotted Flat | <i>Celaenorrhinus leucocera chinensis</i> |
| 302 | Hesperiidae | Dark Yellow-banded Flat | <i>Celaenorrhinus aurivittata aurivittata</i> |
| 303 | Hesperiidae | Himalayan White Flat | <i>Seseria dohertyi</i> |
| 304 | Hesperiidae | Fulvous Pied Flat | <i>Pseudocoladenia dan</i> |
| 305 | Hesperiidae | Hairy Angle | <i>Darpa hanria</i> |
| 306 | Hesperiidae | White Yellow-breast Flat | <i>Gerosis sinica indica</i> |
| 307 | Hesperiidae | Water Snow Flat | <i>Tagiades litigiosa litigiosa</i> |
| 308 | Hesperiidae | Yellow Flat | <i>Mooreana trichoneura pralaya</i> |
| 309 | Hesperiidae | Chestnut Angle | <i>Odontoptilum angulata</i> |
| 310 | Hesperiidae | Tawny Angle | <i>Ctenoptilum vasava vasava</i> |
| 311 | Hesperiidae | Striped Dawnfly | <i>Capila jayadeva</i> |
| 312 | Hesperiidae | Small Indian Palm Bob | <i>Suastus minuta aditia</i> |
| 313 | Hesperiidae | Common Dartlet | <i>Oriens gola gola</i> |
| 314 | Hesperiidae | Common Dart | <i>Potanthus pseudomaesa</i> |
| 315 | Hesperiidae | Broad Bident Dart | <i>Potanthus trachala tytleri</i> |
| 316 | Hesperiidae | Chinese Dart | <i>Potanthus confucius</i> |
| 317 | Hesperiidae | Sikkim Dart | <i>Potanthus mara</i> |
| 318 | Hesperiidae | Pale Palm Dart | <i>Telicota colon</i> |

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| 319 | Hesperiidae | Dark Palm Dart | <i>Telicota ancilla</i> |
| 320 | Hesperiidae | Light Straw Ace | <i>Pithauria stramineipennis</i> |
| 321 | Hesperiidae | Dark Straw Ace | <i>Pithauria murdava</i> |
| 322 | Hesperiidae | Chequered Ace | <i>Thoessa hyrie</i> |
| 323 | Hesperiidae | Ace sp. | <i>Thoessa sp.</i> |
| 324 | Hesperiidae | Northern Spotted Ace | <i>Thoessa cerata</i> |
| 325 | Hesperiidae | Banded Ace | <i>Halpe zema zema</i> |
| 326 | Hesperiidae | Moore's Ace | <i>Halpe porus</i> |
| 327 | Hesperiidae | Plain Ace | <i>Halpe kumara</i> |
| 328 | Hesperiidae | Indian Ace | <i>Halpe homolea</i> |
| 329 | Hesperiidae | Tufted Ace | <i>Sebastonyma dolopia</i> |
| 330 | Hesperiidae | Brown Bush Bob | <i>Pedesta pandita</i> |
| 331 | Hesperiidae | Bush Bob sp | <i>Pedesta sp.</i> |
| 332 | Hesperiidae | Figure of 8 Swift | <i>Caltoris pagana</i> |
| 333 | Hesperiidae | Colon Swift | <i>Caltoris cahira cara</i> |
| 334 | Hesperiidae | Paintbrush Swift | <i>Baoris farri</i> |
| 335 | Hesperiidae | Contiguous Swift | <i>Polytremis lubricans</i> |
| 336 | Hesperiidae | Yellow-Spot Swift | <i>Polytremis eltola</i> |
| 337 | Hesperiidae | Straight Swift | <i>Parnara bada</i> |
| 338 | Hesperiidae | Bevan's Swift | <i>Pseudoborbo bevani</i> |
| 339 | Hesperiidae | Tree Flitter | <i>Hyarotis adrastus praba</i> |
| 340 | Hesperiidae | Purple and Gold Flitter | <i>Zographetus satwa</i> |
| 341 | Hesperiidae | Black-veined Redeye | <i>Matapa sasivarna</i> |
| 342 | Hesperiidae | Hedge Hopper | <i>Baracus vittatus septentrionum</i> |
| 343 | Hesperiidae | Large Forest Bob | <i>Scobura cephaloides cephaloides</i> |
| 344 | Hesperiidae | Dark Velvet Bob | <i>Koruthaialos butleri butleri</i> |
| 345 | Hesperiidae | Chocolate Demon | <i>Ancistroides nigrita</i> |
| 346 | Hesperiidae | Common Banded Demon | <i>Notocrypta paralysos alysia</i> |
| 347 | Hesperiidae | Spotted Demon | <i>Notocrypta feisthameli alysos</i> |
| 348 | Hesperiidae | Grass Demon | <i>Udaspes folus</i> |
| 349 | Hesperiidae | Forest Hopper | <i>Asticopterus jama kada</i> |
| 350 | Hesperiidae | Veined Scrub Hopper | <i>Aeromachus stigmatus</i> |
| 351 | Hesperiidae | Grey Scrub Hopper | <i>Aeromachus jhora creta</i> |
| 352 | Hesperiidae | Hedge Hopper sp. | <i>Baracus sp.</i> |
| 353 | Hesperiidae | Manipur Ace | <i>Sovia malta</i> |
| 354 | Hesperiidae | Lucas' Ace | <i>Sovia magna</i> |

APPENDIX II: INFORMATION ON FISH SPECIES OF DIBANG VALLEY (various sources)

Fish species described from Dibang River basin in the past 10 years:

1. *Aborichthys iphipaniensis* (Kosygin et al., 2019) - Ihipani River at Roing, Lower Dibang Valley, Arunachal Pradesh, India, 28°10'44"N, 95°50'08"E, elevation 418 meters.
2. *Garra arunachalensis* (Nebeshwar and Vishwanath, 2013) - Lower Dibang valley district, Deapani River at Roing, Brahmaputa basin, 29°09'35"N, 95°54'08"E, Arunachal Pradesh, India.
3. *Garra arupi* Nebeshwar (Vishwanath and Das 2009) - Deopani River at Roing, Lower Dibang Valley, Arunachal Pradesh, India.
4. *Mustura harkishorei* (Das and Darshan, 2017) - Dibang River, Brahmaputra basin, Lower Dibang Valley district, Arunachal Pradesh state, India, 28°09'59"N, 95°43'55"E.
5. *Mystus prabini* (Darshan et al., 2019) - Sinkin River (a tributary of the Siang River) at Anpum village, Lower Dibang district, Arunachal Pradesh, India, 28°0'4.4"N, 95°35'9.6"E.
6. *Pseudolaguvia jiyaensis* (Tamang and Sinha, 2014) - Arunachal Pradesh, Lower Dibang Valley District, Jiya stream, near Bolik village, approx. 14 km from Roing towards Shantipur, Assam, India, 28°00.377"N, 95°45.562"E, depth 149 meters.
7. *Pseudolaguvia magna* (Tamang and Sinha, 2014) - Arunachal Pradesh, Lower Dibang Valley District, Jiya stream, near Bolik village, approx. 14 km from Roing towards Shantipur, Assam, India, 28°00.377"N, 95°45.562"E.
8. *Aborichthys waikhomi* (Singh et. al., 2018) - Arunachal Pradesh, Sinkin river at Anpum, Lower Dibang Valley district

Recently recorded fish species from Lower Dibang Valley (Darshan et al., 2019):

- *Anguilla bengalensis* (Gray 1831)
- *Salmostoma phulo* (Hamilton 1822)
- *Garra arupi* Nebeshwar, Waikhom and Das 2009
- *Neolissochilus hexastichus* (McClelland 1839)
- *Systemus immaculatus* McClelland 1839
- *Systemus sarana* (Hamilton 1822)
- *Schizothorax progastus* (McClelland 1839)
- *Cyprinion semiplotum* (McClelland 1839)
- *Botia rostrata* Gunther 1868
- *Lepidocephalichthys arunachalensis* Datta and Barman 1984
- *Aborichthys elongatus* Hora 1921
- *Aborichthys waikhomi* Kosygin 2012
- *Schistura zonata* McClelland 1839
- *Schistura devdevi* Hora 1935
- *Schistura savona* (Hamilton 1822)
- *Mustura harkishorei* Das and Darshan 2017
- *Ompok pabda* (Hamilton 1822)
- *Batasio batasio* (Hamilton 1822)
- *Mystus dibrugarensis* Chaudhuri 1931
- *Amblyceps laticeps* (McClelland 1842)

- *Exostoma labiatum* (McClelland 1842)
- *Glyptothorax cavia* (Hamilton 1822)
- *Pseudecheneis sirenica* Vishwanath and Darshan 2007
- *Pseudolaguvia jiyaensis* Tamang and Sinha 2014
- *Pseudolaguvia magna* Tamang and Sinha 2014
- *Xenentodon cancila* (Hamilton 1822)
- *Macrognaathus pancalus* Hamilton 1822
- *Channa stewartii* (Playfair 1867)
- *Nandus nandus* (Hamilton 1822)
- *Badis assamensis* Ahl 1937
- *Microphis deocata* (Hamilton 1822)
- *Leiodon cutcutia* (Hamilton 1822)

APPENDIX III: CHECKLIST OF BIRDS OF DIBANG VALLEY (Dalvi 2020, unpublished list)

| S.No. | Family | Common Name | Scientific Name | WPA 1972 | IUCN Redlist |
|-------|-------------|-----------------------------|----------------------------------|----------|-----------------|
| 1 | Anatidae | Fulvous Whistling Duck | <i>Dendrocygna bicolor</i> | Sch IV | Least Concern |
| 2 | Anatidae | Lesser Whistling Duck | <i>Dendrocygna javanica</i> | Sch I | Least Concern |
| 3 | Anatidae | Greylag Goose | <i>Anser anser</i> | Sch IV | Least Concern |
| 4 | Anatidae | Gadwall | <i>Mareca strepera</i> | Sch IV | Least Concern |
| 5 | Anatidae | Eurasian Wigeon | <i>Mareca penelope</i> | Sch IV | Least Concern |
| 6 | Anatidae | Indian Spot-billed Duck | <i>Anas poecilorhyncha</i> | Sch IV | Least Concern |
| 7 | Anatidae | Tufted Duck | <i>Aythya fuligula</i> | Sch IV | Least Concern |
| 8 | Phasianidae | Hill Partridge | <i>Arborophila torquola</i> | Sch IV | Least Concern |
| 9 | Phasianidae | Rufous-throated Partridge | <i>Arborophila rufogularis</i> | Sch IV | Least Concern |
| 10 | Phasianidae | White-cheeked Partridge | <i>Arborophila atrogularis</i> | Sch IV | Near Threatened |
| 11 | Phasianidae | Chestnut-breasted Partridge | <i>Arborophila mandellii</i> | Sch IV | Vulnerable |
| 12 | Phasianidae | Blood Pheasant | <i>Ithaginis cruentus</i> | Sch I | Least Concern |
| 13 | Phasianidae | Blyth's Tragopan | <i>Tragopan blythii</i> | Sch I | Vulnerable |
| 14 | Phasianidae | Himalayan Monal | <i>Lophophorus impejanus</i> | Sch I | Least Concern |
| 15 | Phasianidae | Slater's Monal | <i>Lophophorus sclateri</i> | Sch I | Vulnerable |
| 16 | Phasianidae | Red Junglefowl | <i>Gallus gallus</i> | Sch IV | Least Concern |
| 17 | Phasianidae | Kalij Pheasant | <i>Lophura leucomelanos</i> | Sch IV | Least Concern |
| 18 | Phasianidae | Grey Peacock-Pheasant | <i>Polyplectron bicalcaratum</i> | Sch I | Least Concern |
| 19 | Ciconiidae | Asian Openbill | <i>Anastomus oscitans</i> | Sch IV | Least Concern |
| 20 | Ciconiidae | Black Stork | <i>Ciconia nigra</i> | Sch IV | Least Concern |
| 21 | Ciconiidae | Lesser Adjutant | <i>Leptoptilos javanicus</i> | Sch IV | Vulnerable |
| 22 | Ardeidae | Cinnamon Bittern | <i>Ixobrychus cinnamomeus</i> | Sch IV | Least Concern |
| 23 | Ardeidae | Black-crowned Night Heron | <i>Nycticorax nycticorax</i> | Sch IV | Least Concern |
| 24 | Ardeidae | Indian Pond Heron | <i>Ardeola grayii</i> | Sch IV | Least Concern |
| 25 | Ardeidae | Chinese Pond Heron | <i>Ardeola bacchus</i> | Sch IV | Least Concern |
| 26 | Ardeidae | Eastern Cattle Egret | <i>Bubulcus coromandus</i> | Sch IV | Least Concern |
| 27 | Ardeidae | Grey Heron | <i>Ardea cinerea</i> | Sch IV | Least Concern |
| 28 | Ardeidae | Purple Heron | <i>Ardea purpurea</i> | Sch IV | Least Concern |
| 29 | Ardeidae | Great Egret | <i>Ardea alba</i> | Sch IV | Least Concern |
| 30 | Ardeidae | Intermediate Egret | <i>Ardea intermedia</i> | Sch IV | Least Concern |

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|----|-------------------|-------------------------|---------------------------------|--------|-----------------------|
| 31 | Ardeidae | Little Egret | <i>Egretta garzetta</i> | Sch IV | Least Concern |
| 32 | Phalacrocoracidae | Little Cormorant | <i>Microcarbo niger</i> | Sch IV | Least Concern |
| 33 | Phalacrocoracidae | Great Cormorant | <i>Phalacrocorax carbo</i> | Sch IV | Least Concern |
| 34 | Pandionidae | Western Osprey | <i>Pandion haliaetus</i> | Sch I | Least Concern |
| 35 | Accipitridae | White-rumped Vulture | <i>Gyps bengalensis</i> | Sch I | Critically Endangered |
| 36 | Accipitridae | Crested Serpent Eagle | <i>Spilornis cheela</i> | Sch IV | Least Concern |
| 37 | Accipitridae | Short-toed Snake Eagle | <i>Circaetus gallicus</i> | Sch IV | Least Concern |
| 38 | Accipitridae | Mountain Hawk-Eagle | <i>Nisaetus nipalensis</i> | Sch IV | Least Concern |
| 39 | Accipitridae | Rufous-bellied Eagle | <i>Lophotriorchis kienerii</i> | Sch IV | Least Concern |
| 40 | Accipitridae | Black Eagle | <i>Ictinaetus malaiensis</i> | Sch IV | Least Concern |
| 41 | Accipitridae | Greater Spotted Eagle | <i>Clanga clanga</i> | Sch IV | Vulnerable |
| 42 | Accipitridae | Booted Eagle | <i>Hieraaetus pennatus</i> | Sch IV | Least Concern |
| 43 | Accipitridae | Steppe Eagle | <i>Aquila nipalensis</i> | Sch IV | Endangered |
| 44 | Accipitridae | Crested Goshawk | <i>Accipiter trivirgatus</i> | Sch IV | Least Concern |
| 45 | Accipitridae | Shikra | <i>Accipiter badius</i> | Sch IV | Least Concern |
| 46 | Accipitridae | Besra | <i>Accipiter virgatus</i> | Sch IV | Least Concern |
| 47 | Accipitridae | Eurasian Sparrowhawk | <i>Accipiter nisus</i> | Sch IV | Least Concern |
| 48 | Accipitridae | Northern Goshawk | <i>Accipiter gentilis</i> | Sch IV | Least Concern |
| 49 | Accipitridae | Western Marsh Harrier | <i>Circus aeruginosus</i> | Sch IV | Least Concern |
| 50 | Accipitridae | Pallid Harrier | <i>Circus macrourus</i> | Sch IV | Near Threatened |
| 51 | Accipitridae | Pied Harrier | <i>Circus melanoleucos</i> | Sch IV | Least Concern |
| 52 | Accipitridae | Black Kite | <i>Milvus migrans</i> | Sch IV | Least Concern |
| 53 | Accipitridae | Himalayan Buzzard | <i>Buteo burmanicus</i> | Sch IV | Least Concern |
| 54 | Accipitridae | Common Buzzard | <i>Buteo buteo</i> | Sch IV | Least Concern |
| 55 | Otididae | Bengal Florican | <i>Houbaropsis bengalensis</i> | Sch I | Critically Endangered |
| 56 | Rallidae | White-breasted Waterhen | <i>Amaurornis phoenicurus</i> | Sch IV | Least Concern |
| 57 | Rallidae | Common Moorhen | <i>Gallinula chloropus</i> | Sch IV | Least Concern |
| 58 | Charadriidae | Northern Lapwing | <i>Vanellus vanellus</i> | Sch IV | Near Threatened |
| 59 | Charadriidae | Red-wattled Lapwing | <i>Vanellus indicus</i> | Sch IV | Least Concern |
| 60 | Charadriidae | Little Ringed Plover | <i>Charadrius dubius</i> | Sch IV | Least Concern |
| 61 | Charadriidae | Kentish Plover | <i>Charadrius alexandrinus</i> | Sch IV | Least Concern |
| 62 | Jacanidae | Pheasant-tailed Jacana | <i>Hydrophasianus chirurgus</i> | Sch IV | Least Concern |

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|----|--------------|---------------------------------------|---|--------|--------------------|
| 63 | Scolopacidae | Eurasian Woodcock | <i>Scolopax rusticola</i> | Sch IV | Least Concern |
| 64 | Scolopacidae | Solitary Snipe | <i>Gallinago solitaria</i> | Sch IV | Least Concern |
| 65 | Scolopacidae | Wood Snipe | <i>Gallinago nemoricola</i> | Sch IV | Vulnerable |
| 66 | Scolopacidae | Pin-tailed Snipe Common | <i>Gallinago stenura</i> | Sch IV | Least Concern |
| 67 | Scolopacidae | Greenshank | <i>Tringa nebularia</i> | Sch IV | Least Concern |
| 68 | Scolopacidae | Green Sandpiper | <i>Tringa ochropus</i> | Sch IV | Least Concern |
| 69 | Scolopacidae | Wood Sandpiper | <i>Tringa glareola</i> | Sch IV | Least Concern |
| 70 | Scolopacidae | Common Sandpiper | <i>Actitis hypoleucos</i> | Sch IV | Least Concern |
| 71 | Scolopacidae | Temminck's Stint | <i>Calidris temminckii</i> | Sch IV | Least Concern |
| 72 | Glareolidae | Small Pratincole | <i>Glareola lactea</i> | Sch IV | Least Concern |
| 73 | Laridae | Pallas's Gull Speckled Wood | <i>Ichthyaetus ichthyaetus</i> | Sch IV | Least Concern |
| 74 | Columbidae | Pigeon | <i>Columba hodgsonii</i> | Sch IV | Least Concern |
| 75 | Columbidae | Ashy Wood Pigeon | <i>Columba pulchricollis</i> | Sch IV | Least Concern |
| 76 | Columbidae | Pale-capped Pigeon Oriental Turtle | <i>Columba punicea</i> | Sch IV | Vulnerable |
| 77 | Columbidae | Dove | <i>Streptopelia orientalis</i> | Sch IV | Least Concern |
| 78 | Columbidae | Spotted Dove Barred Cuckoo- | <i>Spilopelia chinensis</i> | Sch IV | Least Concern |
| 79 | Columbidae | Dove Common Emerald | <i>Macropygia unchall</i> | Sch IV | Least Concern |
| 80 | Columbidae | Dove Ashy-headed Green | <i>Chalcophaps indica</i> | Sch IV | Least Concern |
| 81 | Columbidae | Pigeon Green Imperial | <i>Treron phayrei</i> | Sch IV | Near Threatened |
| 82 | Columbidae | Pigeon Mountain Imperial | <i>Ducula aenea</i> | Sch IV | Least Concern |
| 83 | Columbidae | Pigeon | <i>Ducula badia</i> | Sch IV | Least Concern |
| 84 | Cuculidae | Greater Coucal | <i>Centropus sinensis</i> | Sch IV | Least Concern |
| 85 | Cuculidae | Lesser Coucal Green-billed | <i>Centropus bengalensis</i> | Sch IV | Least Concern |
| 86 | Cuculidae | Malkoha | <i>Phaenicophaeus tristis</i> <i>Eudynamys</i> | Sch IV | Least Concern |
| 87 | Cuculidae | Asian Koel Asian Emerald | <i>scolopaceus</i> <i>Chrysococcyx</i> | Sch IV | Least Concern |
| 88 | Cuculidae | Cuckoo Banded Bay | <i>maculatus</i> | Sch IV | Least Concern |
| 89 | Cuculidae | Cuckoo | <i>Cacomantis sonneratii</i> | Sch IV | Least Concern |
| 90 | Cuculidae | Plaintive Cuckoo Square-tailed | <i>Cacomantis merulinus</i> | Sch IV | Least Concern |
| 91 | Cuculidae | Drongo-Cuckoo Large Hawk- | <i>Surniculus lugubris</i> <i>Hierococcyx</i> | Sch IV | Least Concern |
| 92 | Cuculidae | Cuckoo Common Hawk- | <i>sparveroides</i> | Sch IV | Least Concern |
| 93 | Cuculidae | Cuckoo Hodgson's Hawk- | <i>Hierococcyx varius</i> | Sch IV | Least Concern |
| 94 | Cuculidae | Cuckoo | <i>Hierococcyx nasicolor</i> | Sch IV | Least Concern |

| | | | | | |
|-----|---------------|---------------------------|----------------------------------|--------|-----------------|
| 95 | Cuculidae | Lesser Cuckoo | <i>Cuculus poliocephalus</i> | Sch IV | Least Concern |
| 96 | Cuculidae | Indian Cuckoo | <i>Cuculus micropterus</i> | Sch IV | Least Concern |
| 97 | Cuculidae | Himalayan Cuckoo | <i>Cuculus saturatus</i> | Sch IV | Least Concern |
| 98 | Cuculidae | Common Cuckoo | <i>Cuculus canorus</i> | Sch IV | Least Concern |
| 99 | Tytonidae | Western Barn Owl | <i>Tyto alba</i> | Sch IV | Least Concern |
| 100 | Strigidae | Mountain Scops Owl | <i>Otus spilocephalus</i> | Sch IV | Least Concern |
| 101 | Strigidae | Collared Scops Owl | <i>Otus lettia</i> | Sch IV | Least Concern |
| 102 | Strigidae | Oriental Scops Owl | <i>Otus sunia</i> | Sch IV | Least Concern |
| 103 | Strigidae | Brown Fish Owl | <i>Ketupa zeylonensis</i> | Sch IV | Least Concern |
| 104 | Strigidae | Brown Wood Owl | <i>Strix leptogrammica</i> | Sch IV | Least Concern |
| 105 | Strigidae | Himalayan Owl | <i>Strix nivicolom</i> | Sch IV | Least Concern |
| 106 | Strigidae | Collared Owlet | <i>Glaucidium brodiei</i> | Sch IV | Least Concern |
| 107 | Strigidae | Asian Barred Owlet | <i>Glaucidium cuculoides</i> | Sch IV | Least Concern |
| 108 | Strigidae | Spotted Owlet | <i>Athene brama</i> | Sch IV | Least Concern |
| 109 | Strigidae | Long-eared Owl | <i>Asio otus</i> | Sch IV | Least Concern |
| 110 | Podargidae | Hodgson's Frogmouth | <i>Batrachostomus hodgsoni</i> | Sch IV | Least Concern |
| 111 | Caprimulgidae | Large-tailed Nightjar | <i>Caprimulgus macrurus</i> | Sch IV | Least Concern |
| 112 | Apodidae | Himalayan Swiftlet | <i>Aerodramus brevirostris</i> | Sch IV | Least Concern |
| 113 | Apodidae | White-throated Needletail | <i>Hirundapus caudacutus</i> | Sch IV | Least Concern |
| 114 | Apodidae | Asian Palm Swift | <i>Cypsiurus balasiensis</i> | Sch IV | Least Concern |
| 115 | Apodidae | Dark-rumped Swift | <i>Apus acuticauda</i> | Sch IV | Vulnerable |
| 116 | Trogonidae | Red-headed Trogon | <i>Harpactes erythrocephalus</i> | Sch IV | Least Concern |
| 117 | Trogonidae | Ward's Trogon | <i>Harpactes wardi</i> | Sch IV | Near Threatened |
| 118 | Alcedinidae | Ruddy Kingfisher | <i>Halcyon coromanda</i> | Sch IV | Least Concern |
| 119 | Alcedinidae | White-throated Kingfisher | <i>Halcyon smyrnensis</i> | Sch IV | Least Concern |
| 120 | Alcedinidae | Blue-eared Kingfisher | <i>Alcedo meninting</i> | Sch IV | Least Concern |
| 121 | Alcedinidae | Crested Kingfisher | <i>Megaceryle lugubris</i> | Sch IV | Least Concern |
| 122 | Alcedinidae | Pied Kingfisher | <i>Ceryle rudis</i> | Sch IV | Least Concern |
| 123 | Meropidae | Blue-bearded Bee-eater | <i>Nyctyornis athertoni</i> | Sch IV | Least Concern |
| 124 | Meropidae | Green Bee-eater | <i>Merops orientalis</i> | Sch IV | Least Concern |
| 125 | Meropidae | Chestnut-headed Bee-eater | <i>Merops leschenaulti</i> | Sch IV | Least Concern |
| 126 | Upupidae | Eurasian Hoopoe | <i>Upupa epops</i> | Sch IV | Least Concern |
| 127 | Bucerotidae | Rufous-necked Hornbill | <i>Aceros nipalensis</i> | Sch I | Vulnerable |

| | | | | | |
|-----|-------------------|------------------------------------|---|--------|--------------------|
| 128 | Bucerotidae | Wreathed Hornbill | <i>Rhyticeros undulatus</i> | Sch I | Least Concern |
| 129 | Megalaimidae | Great Barbet | <i>Psilopogon virens</i> | Sch IV | Least Concern |
| 130 | Megalaimidae | Lineated Barbet Golden-throated | <i>Psilopogon lineatus</i> | Sch IV | Least Concern |
| 131 | Megalaimidae | Barbet Blue-throated | <i>Psilopogon franklinii</i> | Sch IV | Least Concern |
| 132 | Megalaimidae | Barbet | <i>Psilopogon asiaticus</i> | Sch IV | Least Concern |
| 133 | Megalaimidae | Blue-eared Barbet | <i>Psilopogon duvaucelii</i> | Sch IV | Least Concern |
| 134 | Picidae | Speckled Piculet White-browed | <i>Picumnus innominatus</i> | Sch IV | Least Concern |
| 135 | Picidae | Piculet Rufous-bellied | <i>Sasia ochracea</i> <i>Dendrocopos</i> | Sch IV | Least Concern |
| 136 | Picidae | Woodpecker Fulvous-breasted | <i>hyperythrus</i> | Sch IV | Least Concern |
| 137 | Picidae | Woodpecker Crimson-breasted | <i>Dendrocopos macei</i> | Sch IV | Least Concern |
| 138 | Picidae | Woodpecker Darjeeling | <i>Dryobates cathpharius</i> <i>Dendrocopos</i> | Sch IV | Least Concern |
| 139 | Picidae | Woodpecker | <i>darjellensis</i> <i>Chrysophlegma</i> | Sch IV | Least Concern |
| 140 | Picidae | Greater Yellownappe | <i>flavinucha</i> | Sch IV | Least Concern |
| 141 | Picidae | Lesser Yellownappe Grey-headed | <i>Picus chlorolophus</i> | Sch IV | Least Concern |
| 142 | Picidae | Woodpecker Pale-headed | <i>Picus canus</i> | Sch IV | Least Concern |
| 143 | Picidae | Woodpecker | <i>Gecinulus grantia</i> | Sch IV | Least Concern |
| 144 | Picidae | Bay Woodpecker | <i>Blythipicus pyrrhotis</i> <i>Micropternus</i> | Sch IV | Least Concern |
| 145 | Picidae | Rufous Woodpecker Great Slaty | <i>brachyurus</i> <i>Mulleripicus</i> | Sch IV | Least Concern |
| 146 | Picidae | Woodpecker | <i>pulverulentus</i> | Sch IV | Vulnerable |
| 147 | Falconidae | Amur Falcon | <i>Falco amurensis</i> | Sch IV | Least Concern |
| 148 | Falconidae | Eurasian Hobby | <i>Falco subbuteo</i> | Sch IV | Least Concern |
| 149 | Falconidae | Oriental Hobby | <i>Falco severus</i> | Sch IV | Least Concern |
| 150 | Falconidae | Peregrine Falcon Grey-headed | <i>Falco peregrinus</i> | Sch I | Least Concern |
| 151 | Psittaculidae | Parakeet Red-breasted | <i>Psittacula finschii</i> | Sch IV | Near Threatened |
| 152 | Psittaculidae | Parakeet Rose-ringed | <i>Psittacula alexandri</i> | Sch IV | Near Threatened |
| 153 | Psittaculidae | Parakeet Long-tailed | <i>Psittacula krameri</i> | Sch IV | Least Concern |
| 154 | Eurylaimidae | Broadbill Silver-breasted | <i>Psarisomus dalhousiae</i> | Sch IV | Least Concern |
| 155 | Eurylaimidae | Broadbill | <i>Serilophus lunatus</i> | Sch IV | Least Concern |
| 156 | Pittidae | Hooded Pitta Bar-winged | <i>Pitta sordida</i> | Sch IV | Least Concern |
| 157 | Tephrodornithidae | Flycatcher-shrike | <i>Hemipus picatus</i> | Sch IV | Least Concern |
| 158 | Tephrodornithidae | Large Woodshrike | <i>Tephrodornis virgatus</i> | Sch IV | Least Concern |

| | | | | | |
|-----|---------------|----------------------------------|--|--------|---------------|
| 159 | Aegithinidae | Common Iora | <i>Aegithina tiphia</i> | Sch IV | Least Concern |
| 160 | Campephagidae | Large Cuckooshrike | <i>Coracina macei</i> | Sch IV | Least Concern |
| 161 | Campephagidae | Black-winged Cuckooshrike | <i>Lalage melaschistos</i> | Sch IV | Least Concern |
| 162 | Campephagidae | Grey-chinned Minivet | <i>Pericrocotus solaris</i> | Sch IV | Least Concern |
| 163 | Campephagidae | Long-tailed Minivet | <i>Pericrocotus ethologus</i> <i>Pericrocotus</i> | Sch IV | Least Concern |
| 164 | Campephagidae | Short-billed Minivet | <i>brevirostris</i> | Sch IV | Least Concern |
| 165 | Campephagidae | Scarlet Minivet | <i>Pericrocotus speciosus</i> | Sch IV | Least Concern |
| 166 | Laniidae | Brown Shrike | <i>Lanius cristatus</i> | Sch IV | Least Concern |
| 167 | Laniidae | Burmese Shrike | <i>Lanius colluriooides</i> | Sch IV | Least Concern |
| 168 | Laniidae | Long-tailed Shrike | <i>Lanius schach</i> | Sch IV | Least Concern |
| 169 | Laniidae | Grey-backed Shrike | <i>Lanius tephronotus</i> | Sch IV | Least Concern |
| 170 | Vireonidae | Black-headed Shrike-Babbler | <i>Pteruthius rufiventer</i> | Sch IV | Least Concern |
| 171 | Vireonidae | Green Shrike- Babbler | <i>Pteruthius</i> <i>xanthochlorus</i> | Sch IV | Least Concern |
| 172 | Vireonidae | Black-eared Shrike- Babbler | <i>Pteruthius melanotis</i> | Sch IV | Least Concern |
| 173 | Oriolidae | Black-hooded Oriole | <i>Oriolus xanthornus</i> | Sch IV | Least Concern |
| 174 | Oriolidae | Maroon Oriole | <i>Oriolus traillii</i> | Sch IV | Least Concern |
| 175 | Dicruridae | Black Drongo | <i>Dicrurus macrocercus</i> | Sch IV | Least Concern |
| 176 | Dicruridae | Ashy Drongo | <i>Dicrurus leucophaeus</i> | Sch IV | Least Concern |
| 177 | Dicruridae | Bronzed Drongo | <i>Dicrurus aeneus</i> | Sch IV | Least Concern |
| 178 | Dicruridae | Lesser Racket-tailed Drongo | <i>Dicrurus remifer</i> | Sch IV | Least Concern |
| 179 | Dicruridae | Greater Racket- tailed Drongo | <i>Dicrurus paradiseus</i> | Sch IV | Least Concern |
| 180 | Rhipiduridae | White-throated Fantail | <i>Rhipidura albicollis</i> | Sch IV | Least Concern |
| 181 | Monarchidae | Black-naped Monarch | <i>Hypothymis azurea</i> | Sch IV | Least Concern |
| 182 | Monarchidae | Blyth's Paradise Flycatcher | <i>Terpsiphone affinis</i> | Sch IV | Least Concern |
| 183 | Corvidae | Eurasian Jay | <i>Garrulus glandarius</i> | Sch IV | Least Concern |
| 184 | Corvidae | Common Green Magpie | <i>Cissa chinensis</i> | Sch IV | Least Concern |
| 185 | Corvidae | Rufous Treepie | <i>Dendrocitta vagabunda</i> | Sch IV | Least Concern |
| 186 | Corvidae | Grey Treepie | <i>Dendrocitta formosae</i> | Sch IV | Least Concern |
| 187 | Corvidae | Collared Treepie | <i>Dendrocitta frontalis</i> | Sch IV | Least Concern |
| 188 | Corvidae | House Crow | <i>Corvus splendens</i> | Sch IV | Least Concern |
| 189 | Corvidae | Large-billed Crow | <i>Corvus macrorhynchos</i> | Sch IV | Least Concern |
| 190 | Stenostiridae | Grey-headed Canary-flycatcher | <i>Culicicapa ceylonensis</i> | Sch IV | Least Concern |
| 191 | Paridae | Yellow-browed Tit | <i>Sylviparus modestus</i> | Sch IV | Least Concern |

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|-----|----------------|---------------------------|--------------------------------------|--------|---------------|
| 192 | Paridae | Sultan Tit | <i>Melanochlora sultanea</i> | Sch IV | Least Concern |
| 193 | Paridae | Rufous-vented Tit | <i>Periparus rubidiventris</i> | Sch IV | Least Concern |
| 194 | Paridae | Coal Tit | <i>Periparus ater</i> | Sch IV | Least Concern |
| 195 | Paridae | Grey Crested Tit | <i>Lophophanes dichrous</i> | Sch IV | Least Concern |
| 196 | Paridae | Green-backed Tit | <i>Parus monticolus</i> | Sch IV | Least Concern |
| 197 | Paridae | Yellow-cheeked Tit | <i>Machlolophus silonotus</i> | Sch IV | Least Concern |
| 198 | Alaudidae | Bengal Bush Lark | <i>Mirafra assamica</i> | Sch IV | Least Concern |
| 199 | Alaudidae | Oriental Skylark | <i>Alauda gulgula</i> | Sch IV | Least Concern |
| 200 | Pycnonotidae | Striated Bulbul | <i>Pycnonotus striatus</i> | Sch IV | Least Concern |
| 201 | Pycnonotidae | Black-crested Bulbul | <i>Pycnonotus flaviventris</i> | Sch IV | Least Concern |
| 202 | Pycnonotidae | Red-whiskered Bulbul | <i>Pycnonotus jocosus</i> | Sch IV | Least Concern |
| 203 | Pycnonotidae | Red-vented Bulbul | <i>Pycnonotus cafer</i> | Sch IV | Least Concern |
| 204 | Pycnonotidae | White-throated Bulbul | <i>Alophoixus flaveolus</i> | Sch IV | Least Concern |
| 205 | Pycnonotidae | Mountain Bulbul | <i>Ixos mcclellandii</i> | Sch IV | Least Concern |
| 206 | Pycnonotidae | Ashy Bulbul | <i>Hemixos flavala</i> | Sch IV | Least Concern |
| 207 | Hirundinidae | Barn Swallow | <i>Hirundo rustica</i> | Sch IV | Least Concern |
| 208 | Hirundinidae | Asian House Martin | <i>Delichon dasypus</i> | Sch IV | Least Concern |
| 209 | Hirundinidae | Nepal House Martin | <i>Delichon nipalense</i> | Sch IV | Least Concern |
| 210 | Hirundinidae | Striated Swallow | <i>Cecropis striolata</i> | Sch IV | Least Concern |
| 211 | Cettiidae | Yellow-bellied Warbler | <i>Abroscopus superciliaris</i> | Sch IV | Least Concern |
| 212 | Cettiidae | Rufous-faced Warbler | <i>Abroscopus albogularis</i> | Sch IV | Least Concern |
| 213 | Cettiidae | Black-faced Warbler | <i>Abroscopus schisticeps</i> | Sch IV | Least Concern |
| 214 | Cettiidae | Mountain Tailorbird | <i>Phyllergates cucullatus</i> | Sch IV | Least Concern |
| 215 | Cettiidae | Broad-billed Warbler | <i>Tickellia hodgsoni</i> | Sch IV | Least Concern |
| 216 | Cettiidae | Grey-bellied Tesia | <i>Tesia cyaniventer</i> | Sch IV | Least Concern |
| 217 | Cettiidae | Slaty-bellied Tesia | <i>Tesia olivea</i> | Sch IV | Least Concern |
| 218 | Cettiidae | Chestnut-headed Tesia | <i>Cettia castaneocoronata</i> | Sch IV | Least Concern |
| 219 | Phylloscopidae | Dusky Warbler | <i>Phylloscopus fuscatus</i> | Sch IV | Least Concern |
| 220 | Phylloscopidae | Tickell's Leaf Warbler | <i>Phylloscopus affinis</i> | Sch IV | Least Concern |
| 221 | Phylloscopidae | Buff-barred Warbler | <i>Phylloscopus pulcher</i> | Sch IV | Least Concern |
| 222 | Phylloscopidae | Ashy-throated Warbler | <i>Phylloscopus maculipennis</i> | Sch IV | Least Concern |
| 223 | Phylloscopidae | Yellow-browed Warbler | <i>Phylloscopus inornatus</i> | Sch IV | Least Concern |
| 224 | Phylloscopidae | Greenish Warbler | <i>Phylloscopus trochiloides</i> | Sch IV | Least Concern |

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|-----|----------------|----------------------------------|------------------------------------|--------|---------------|
| 225 | Phylloscopidae | Large-billed Leaf Warbler | <i>Phylloscopus magnirostris</i> | Sch IV | Least Concern |
| 226 | Phylloscopidae | Blyth's Leaf Warbler | <i>Phylloscopus reguloides</i> | Sch IV | Least Concern |
| 227 | Phylloscopidae | Yellow-vented Warbler | <i>Phylloscopus cantator</i> | Sch IV | Least Concern |
| 228 | Phylloscopidae | Grey-hooded Warbler | <i>Phylloscopus xanthoschistos</i> | Sch IV | Least Concern |
| 229 | Phylloscopidae | White-spectacled Warbler | <i>Phylloscopus intermedius</i> | Sch IV | Least Concern |
| 230 | Phylloscopidae | Grey-crowned Warbler | <i>Phylloscopus tephrocephalus</i> | Sch IV | Least Concern |
| 231 | Phylloscopidae | Whistler's Warbler | <i>Phylloscopus whistleri</i> | Sch IV | Least Concern |
| 232 | Phylloscopidae | Grey-cheeked Warbler | <i>Phylloscopus poliogenys</i> | Sch IV | Least Concern |
| 233 | Phylloscopidae | Chestnut-crowned Warbler | <i>Phylloscopus castaniceps</i> | Sch IV | Least Concern |
| 234 | Acrocephalidae | Paddyfield Warbler | <i>Acrocephalus agricola</i> | Sch IV | Least Concern |
| 235 | Acrocephalidae | Thick-billed Warbler | <i>Arundinax aedon</i> | Sch IV | Least Concern |
| 236 | Locustellidae | Striated Grassbird | <i>Megalurus palustris</i> | Sch IV | Least Concern |
| 237 | Cisticolidae | Zitting Cisticola | <i>Cisticola juncidis</i> | Sch IV | Least Concern |
| 238 | Cisticolidae | Striated Prinia | <i>Prinia crinigera</i> | Sch IV | Least Concern |
| 239 | Cisticolidae | Black-throated Prinia | <i>Prinia atrogularis</i> | Sch IV | Least Concern |
| 240 | Cisticolidae | Grey-breasted Prinia | <i>Prinia hodgsonii</i> | Sch IV | Least Concern |
| 241 | Cisticolidae | Graceful Prinia | <i>Prinia gracilis</i> | Sch IV | Least Concern |
| 242 | Cisticolidae | Yellow-bellied Prinia | <i>Prinia flaviventris</i> | Sch IV | Least Concern |
| 243 | Cisticolidae | Ashy Prinia | <i>Prinia socialis</i> | Sch IV | Least Concern |
| 244 | Cisticolidae | Plain Prinia | <i>Prinia inornata</i> | Sch IV | Least Concern |
| 245 | Cisticolidae | Common Tailorbird | <i>Orthotomus sutorius</i> | Sch IV | Least Concern |
| 246 | Timaliidae | Spot-breasted Scimitar Babbler | <i>Pomatorhinus mcclellandi</i> | Sch IV | Least Concern |
| 247 | Timaliidae | White-browed Scimitar Babbler | <i>Pomatorhinus schisticeps</i> | Sch IV | Least Concern |
| 248 | Timaliidae | Streak-breasted Scimitar Babbler | <i>Pomatorhinus ruficollis</i> | Sch IV | Least Concern |
| 249 | Timaliidae | Red-billed Scimitar Babbler | <i>Pomatorhinus ochraceiceps</i> | Sch IV | Least Concern |
| 250 | Timaliidae | Coral-billed Scimitar Babbler | <i>Pomatorhinus ferruginosus</i> | Sch IV | Least Concern |
| 251 | Timaliidae | Slender-billed Scimitar Babbler | <i>Pomatorhinus superciliaris</i> | Sch IV | Least Concern |
| 252 | Timaliidae | Bar-winged Wren-Babbler | <i>Spelaeornis troglodytoides</i> | Sch IV | Least Concern |
| 253 | Timaliidae | Grey-throated Babbler | <i>Stachyris nigriceps</i> | Sch IV | Least Concern |
| 254 | Timaliidae | Buff-chested Babbler | <i>Stachyridopsis ambigua</i> | Sch IV | Least Concern |

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|-----|----------------|-------------------------------------|---|--------|---------------|
| 255 | Timaliidae | Rufous-capped Babbler | <i>Stachyridopsis ruficeps</i> <i>Stachyridopsis</i> | Sch IV | Least Concern |
| 256 | Timaliidae | Golden Babbler | <i>chrysaea</i> | Sch IV | Least Concern |
| 257 | Timaliidae | Chestnut-capped Babbler | <i>Timalia pileata</i> | Sch IV | Least Concern |
| 258 | Timaliidae | Pin-striped Tit- Babbler | <i>Macronus gularis</i> | Sch IV | Least Concern |
| 259 | Pellorneidae | Yellow-throated Fulvetta | <i>Alcippe cinerea</i> | Sch IV | Least Concern |
| 260 | Pellorneidae | Rufous-winged Fulvetta | <i>Alcippe castaneiceps</i> | Sch IV | Least Concern |
| 261 | Pellorneidae | Rufous-throated Fulvetta | <i>Alcippe rufogularis</i> | Sch IV | Least Concern |
| 262 | Pellorneidae | Brown-cheeked Fulvetta | <i>Alcippe poioicephala</i> | Sch IV | Least Concern |
| 263 | Pellorneidae | Nepal Fulvetta | <i>Alcippe nipalensis</i> | Sch IV | Least Concern |
| 264 | Pellorneidae | Streaked Wren- Babbler | <i>Napothera</i> <i>brevicaudata</i> | Sch IV | Least Concern |
| 265 | Pellorneidae | Eyebrowed Wren- Babbler | <i>Napothera epilepidota</i> | Sch IV | Least Concern |
| 266 | Pellorneidae | Long-billed Wren- Babbler | <i>Rimator malacoptilus</i> | Sch IV | Least Concern |
| 267 | Pellorneidae | White-hooded Babbler | <i>Gampsorhynchus</i> <i>rufulus</i> | Sch IV | Least Concern |
| 268 | Pellorneidae | Spot-throated Babbler | <i>Pellorneum albiventre</i> | Sch IV | Least Concern |
| 269 | Pellorneidae | Marsh Babbler | <i>Pellorneum palustre</i> | Sch IV | Vulnerable |
| 270 | Pellorneidae | Puff-throated Babbler | <i>Pellorneum ruficeps</i> | Sch IV | Least Concern |
| 271 | Pellorneidae | Buff-breasted Babbler | <i>Pellorneum tickelli</i> | Sch IV | Least Concern |
| 272 | Leiothrichidae | Striated Babbler | <i>Argya earlei</i> | Sch IV | Least Concern |
| 273 | Leiothrichidae | White-crested Laughingthrush | <i>Garrulax leucolophus</i> | Sch IV | Least Concern |
| 274 | Leiothrichidae | Rufous-chinned Laughingthrush | <i>Ianthocincla</i> <i>rufogularis</i> | Sch IV | Least Concern |
| 275 | Leiothrichidae | Spotted Laughingthrush | <i>Ianthocincla ocellata</i> | Sch IV | Least Concern |
| 276 | Leiothrichidae | Lesser Necklaced Laughingthrush | <i>Garrulax monileger</i> | Sch IV | Least Concern |
| 277 | Leiothrichidae | Greater Necklaced Laughingthrush | <i>Pterorhinus pectoralis</i> | Sch IV | Least Concern |
| 278 | Leiothrichidae | Rufous-necked Laughingthrush | <i>Pterorhinus ruficollis</i> | Sch IV | Least Concern |
| 279 | Leiothrichidae | Grey-sided Laughingthrush | <i>Pterorhinus caerulatus</i> | Sch IV | Least Concern |
| 280 | Leiothrichidae | Spot-breasted Laughingthrush | <i>Garrulax merulinus</i> | Sch IV | Least Concern |
| 281 | Leiothrichidae | Striated Laughingthrush | <i>Grammatoptila striata</i> | Sch IV | Least Concern |
| 282 | Leiothrichidae | Blue-winged Laughingthrush | <i>Trochalopteron</i> <i>squamatum</i> | Sch IV | Least Concern |

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| 283 | Leiothrichidae | Scaly Laughingthrush Black-faced | <i>Trochalopteron subunicolor</i> | Sch IV | Least Concern |
| 284 | Leiothrichidae | Laughingthrush Chestnut-crowned | <i>Trochalopteron affine</i> | Sch IV | Least Concern |
| 285 | Leiothrichidae | Laughingthrush Assam | <i>Trochalopteron erythrocephalum</i> | Sch IV | Least Concern |
| 286 | Leiothrichidae | Laughingthrush Elliot's | <i>Trochalopteron chrysopterum</i> | Sch IV | Least Concern |
| 287 | Leiothrichidae | Laughingthrush | <i>Trochalopteron elliotii</i> | Sch IV | Least Concern |
| 288 | Leiothrichidae | Himalayan Cutia | <i>Cutia nipalensis</i> | Sch IV | Least Concern |
| 289 | Leiothrichidae | Blue-winged Minla | <i>Actinodura cyanouroptera</i> | Sch IV | Least Concern |
| 290 | Leiothrichidae | Red-tailed Minla | <i>Minla ignotincta</i> | Sch IV | Least Concern |
| 291 | Leiothrichidae | Red-faced Liocichla Rusty-fronted | <i>Liocichla phoenicea</i> | Sch IV | Least Concern |
| 292 | Leiothrichidae | Barwing Streak-throated | <i>Actinodura egertoni</i> | Sch IV | Least Concern |
| 293 | Leiothrichidae | Barwing | <i>Actinodura waldeni</i> | Sch IV | Least Concern |
| 294 | Leiothrichidae | Red-billed Leiothrix Rufous-backed | <i>Leiothrix lutea</i> | Sch IV | Least Concern |
| 295 | Leiothrichidae | Sibia | <i>Leioptila annectens</i> | Sch IV | Least Concern |
| 296 | Leiothrichidae | Grey Sibia | <i>Heterophasia gracilis</i> | Sch IV | Least Concern |
| 297 | Leiothrichidae | Beautiful Sibia | <i>Heterophasia pulchella</i> | Sch IV | Least Concern |
| 298 | Leiothrichidae | Long-tailed Sibia | <i>Heterophasia picaoides</i> | Sch IV | Least Concern |
| 299 | Sylviidae | Fire-tailed Myzornis Golden-breasted | <i>Myzornis pyrrhoura</i> | Sch IV | Least Concern |
| 300 | Sylviidae | Fulvetta | <i>Lioparus chrysotis</i> | Sch IV | Least Concern |
| 301 | Sylviidae | Jerdon's Babbler | <i>Chrysomma altirostre</i> | Sch IV | Vulnerable |
| 302 | Sylviidae | Brown Parrotbill | <i>Cholornis unicolor</i> | Sch IV | Least Concern |
| 303 | Sylviidae | Fulvous Parrotbill Black-throated | <i>Suthora fulvifrons</i> | Sch IV | Least Concern |
| 304 | Sylviidae | Parrotbill Rufous-headed | <i>Suthora nipalensis</i> | Sch IV | Least Concern |
| 305 | Sylviidae | Parrotbill Grey-headed | <i>Psittiparus bakeri</i> | Sch IV | Least Concern |
| 306 | Sylviidae | Parrotbill Black-breasted | <i>Psittiparus gularis</i> | Sch IV | Least Concern |
| 307 | Sylviidae | Parrotbill Pale-billed | <i>Paradoxornis flavirostris</i> | Sch IV | Vulnerable |
| 308 | Sylviidae | Parrotbill | <i>Chleuasicus atrosuperciliaris</i> | Sch IV | Least Concern |
| 309 | Zosteropidae | Striated Yuhina White-naped | <i>Yuhina castaniceps</i> | Sch IV | Least Concern |
| 310 | Zosteropidae | Yuhina | <i>Yuhina bakeri</i> | Sch IV | Least Concern |
| 311 | Zosteropidae | Whiskered Yuhina Stripe-throated | <i>Yuhina flavicollis</i> | Sch IV | Least Concern |
| 312 | Zosteropidae | Yuhina Rufous-vented | <i>Yuhina gularis</i> | Sch IV | Least Concern |
| 313 | Zosteropidae | Yuhina | <i>Yuhina occipitalis</i> | Sch IV | Least Concern |

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| 314 | Zosteropidae | Black-chinned Yuhina | <i>Yuhina nigrimenta</i> | Sch IV | Least Concern |
| 315 | Irenidae | Asian Fairy-bluebird | <i>Irena puella</i> | Sch IV | Least Concern |
| 316 | Sittidae | Chestnut-vented Nuthatch | <i>Sitta nagaensis</i> | Sch IV | Least Concern |
| 317 | Sittidae | Chestnut-bellied Nuthatch | <i>Sitta cinnamoventris</i> | Sch IV | Least Concern |
| 318 | Sittidae | White-tailed Nuthatch | <i>Sitta himalayensis</i> | Sch IV | Least Concern |
| 319 | Sittidae | Velvet-fronted Nuthatch | <i>Sitta frontalis</i> | Sch IV | Least Concern |
| 320 | Sittidae | Beautiful Nuthatch | <i>Sitta formosa</i> | Sch IV | Vulnerable |
| 321 | Certhiidae | Rusty-flanked Treecreeper | <i>Certhia nipalensis</i> | Sch IV | Least Concern |
| 322 | Sturnidae | Spot-winged Starling | <i>Saroglossa spilopterus</i> | Sch IV | Least Concern |
| 323 | Sturnidae | Common Hill Myna | <i>Gracula religiosa</i> | Sch I | Least Concern |
| 324 | Sturnidae | Great Myna | <i>Acridotheres grandis</i> | Sch IV | Least Concern |
| 325 | Sturnidae | Jungle Myna | <i>Acridotheres fuscus</i> | Sch IV | Least Concern |
| 326 | Sturnidae | Common Myna | <i>Acridotheres tristis</i> | Sch IV | Least Concern |
| 327 | Sturnidae | Chestnut-tailed Starling | <i>Sturnia malabarica</i> | Sch IV | Least Concern |
| 328 | Turdidae | Orange-headed Thrush | <i>Geokichla citrina</i> | Sch IV | Least Concern |
| 329 | Turdidae | Long-tailed Thrush | <i>Zoothera dixonii</i> | Sch IV | Least Concern |
| 330 | Turdidae | Long-billed Thrush | <i>Zoothera monticola</i> | Sch IV | Least Concern |
| 331 | Turdidae | Black-breasted Thrush | <i>Turdus dissimilis</i> | Sch IV | Least Concern |
| 332 | Turdidae | White-collared Blackbird | <i>Turdus albocinctus</i> | Sch IV | Least Concern |
| 333 | Turdidae | Grey-winged Blackbird | <i>Turdus boulboul</i> | Sch IV | Least Concern |
| 334 | Turdidae | Chestnut Thrush | <i>Turdus rubrocanus</i> | Sch IV | Least Concern |
| 335 | Turdidae | Grey-sided Thrush | <i>Turdus feae</i> | Sch IV | Vulnerable |
| 336 | Turdidae | Eyebrowed Thrush | <i>Turdus obscurus</i> | Sch IV | Least Concern |
| 337 | Turdidae | Black-throated Thrush | <i>Turdus atrogularis</i> | Sch IV | Least Concern |
| 338 | Turdidae | Red-throated Thrush | <i>Turdus ruficollis</i> | Sch IV | Least Concern |
| 339 | Turdidae | Dusky Thrush | <i>Turdus eunomus</i> | Sch IV | Least Concern |
| 340 | Turdidae | Purple Cochoa | <i>Cochoa purpurea</i> | Sch IV | Least Concern |
| 341 | Turdidae | Green Cochoa | <i>Cochoa viridis</i> | Sch IV | Least Concern |
| 342 | Muscicapidae | Oriental Magpie-Robin | <i>Copsychus saularis</i> | Sch IV | Least Concern |
| 343 | Muscicapidae | White-rumped Shama | <i>Copsychus malabaricus</i> | Sch IV | Least Concern |
| 344 | Muscicapidae | Dark-sided Flycatcher | <i>Muscicapa sibirica</i> | Sch IV | Least Concern |

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|-----|--------------|----------------------------|--------------------------------|--------|-----------------|
| 345 | Muscicapidae | Ferruginous Flycatcher | <i>Muscicapa ferruginea</i> | Sch IV | Least Concern |
| 346 | Muscicapidae | White-gorgeted Flycatcher | <i>Anthipes monileger</i> | Sch IV | Least Concern |
| 347 | Muscicapidae | Rufous-bellied Niltava | <i>Niltava sundara</i> | Sch IV | Least Concern |
| 348 | Muscicapidae | Vivid Niltava | <i>Niltava vivida</i> | Sch IV | Least Concern |
| 349 | Muscicapidae | Large Niltava | <i>Niltava grandis</i> | Sch IV | Least Concern |
| 350 | Muscicapidae | Small Niltava | <i>Niltava macgrigoriae</i> | Sch IV | Least Concern |
| 351 | Muscicapidae | Verditer Flycatcher | <i>Eumyias thalassinus</i> | Sch IV | Least Concern |
| 352 | Muscicapidae | Gould's Shortwing | <i>Heteroxenicus stellatus</i> | Sch IV | Least Concern |
| 353 | Muscicapidae | Rusty-bellied Shortwing | <i>Brachypteryx hyperythra</i> | Sch IV | Near Threatened |
| 354 | Muscicapidae | Lesser Shortwing | <i>Brachypteryx leucophris</i> | Sch IV | Least Concern |
| 355 | Muscicapidae | Indian Blue Robin | <i>Larvivora brunnea</i> | Sch IV | Least Concern |
| 356 | Muscicapidae | Bluethroat | <i>Luscinia svecica</i> | Sch IV | Least Concern |
| 357 | Muscicapidae | Siberian Rubythroat | <i>Calliope calliope</i> | Sch IV | Least Concern |
| 358 | Muscicapidae | White-tailed Robin | <i>Myiomela leucura</i> | Sch IV | Least Concern |
| 359 | Muscicapidae | Rufous-breasted Bush Robin | <i>Tarsiger hyperythrus</i> | Sch IV | Least Concern |
| 360 | Muscicapidae | White-browed Bush Robin | <i>Tarsiger indicus</i> | Sch IV | Least Concern |
| 361 | Muscicapidae | Golden Bush Robin | <i>Tarsiger chrysaeus</i> | Sch IV | Least Concern |
| 362 | Muscicapidae | Little Forktail | <i>Enicurus scouleri</i> | Sch IV | Least Concern |
| 363 | Muscicapidae | Black-backed Forktail | <i>Enicurus immaculatus</i> | Sch IV | Least Concern |
| 364 | Muscicapidae | Slaty-backed Forktail | <i>Enicurus schistaceus</i> | Sch IV | Least Concern |
| 365 | Muscicapidae | White-crowned Forktail | <i>Enicurus leschenaulti</i> | Sch IV | Least Concern |
| 366 | Muscicapidae | Spotted Forktail | <i>Enicurus maculatus</i> | Sch IV | Least Concern |
| 367 | Muscicapidae | Blue Whistling Thrush | <i>Myophonus caeruleus</i> | Sch IV | Least Concern |
| 368 | Muscicapidae | Blue-fronted Robin | <i>Cinclidium frontale</i> | Sch IV | Least Concern |
| 369 | Muscicapidae | Slaty-backed Flycatcher | <i>Ficedula hodgsonii</i> | Sch IV | Least Concern |
| 370 | Muscicapidae | Rufous-gorgeted Flycatcher | <i>Ficedula strophciata</i> | Sch IV | Least Concern |
| 371 | Muscicapidae | Snowy-browed Flycatcher | <i>Ficedula hyperythra</i> | Sch IV | Least Concern |
| 372 | Muscicapidae | Little Pied Flycatcher | <i>Ficedula westermanni</i> | Sch IV | Least Concern |
| 373 | Muscicapidae | Ultramarine Flycatcher | <i>Ficedula superciliaris</i> | Sch IV | Least Concern |
| 374 | Muscicapidae | Slaty-blue Flycatcher | <i>Ficedula tricolor</i> | Sch IV | Least Concern |
| 375 | Muscicapidae | Sapphire Flycatcher | <i>Ficedula sapphira</i> | Sch IV | Least Concern |

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|-----|---------------|------------------------------|----------------------------------|--------|---------------|
| 376 | Muscicapidae | Black Redstart | <i>Phoenicurus ochruros</i> | Sch IV | Least Concern |
| 377 | Muscicapidae | Hodgson's Redstart | <i>Phoenicurus hodgsoni</i> | Sch IV | Least Concern |
| 378 | Muscicapidae | Daurian Redstart | <i>Phoenicurus aureoreus</i> | Sch IV | Least Concern |
| 379 | Muscicapidae | Blue-fronted Redstart | <i>Phoenicurus frontalis</i> | Sch IV | Least Concern |
| 380 | Muscicapidae | Plumbeous Water Redstart | <i>Phoenicurus fuliginosus</i> | Sch IV | Least Concern |
| 381 | Muscicapidae | White-capped Redstart | <i>Phoenicurus leucocephalus</i> | Sch IV | Least Concern |
| 382 | Muscicapidae | Blue Rock Thrush | <i>Monticola solitarius</i> | Sch IV | Least Concern |
| 383 | Muscicapidae | Chestnut-bellied Rock Thrush | <i>Monticola rufiventris</i> | Sch IV | Least Concern |
| 384 | Muscicapidae | Pied Bush Chat | <i>Saxicola caprata</i> | Sch IV | Least Concern |
| 385 | Muscicapidae | Grey Bush Chat | <i>Saxicola ferreus</i> | Sch IV | Least Concern |
| 386 | Muscicapidae | Isabelline Wheatear | <i>Oenanthe isabellina</i> | Sch IV | Least Concern |
| 387 | Cinclidae | Brown Dipper | <i>Cinclus pallasii</i> | Sch IV | Least Concern |
| 388 | Dicaeidae | Yellow-bellied Flowerpecker | <i>Dicaeum melanoxanthum</i> | Sch IV | Least Concern |
| 389 | Dicaeidae | Plain Flowerpecker | <i>Dicaeum minullum</i> | Sch IV | Least Concern |
| 390 | Dicaeidae | Fire-breasted Flowerpecker | <i>Dicaeum ignipectus</i> | Sch IV | Least Concern |
| 391 | Dicaeidae | Scarlet-backed Flowerpecker | <i>Dicaeum cruentatum</i> | Sch IV | Least Concern |
| 392 | Nectariniidae | Ruby-cheeked Sunbird | <i>Chalcoparia singalensis</i> | Sch IV | Least Concern |
| 393 | Nectariniidae | Purple Sunbird | <i>Cinnyris asiaticus</i> | Sch IV | Least Concern |
| 394 | Nectariniidae | Green-tailed Sunbird | <i>Aethopyga nipalensis</i> | Sch IV | Least Concern |
| 395 | Nectariniidae | Black-throated Sunbird | <i>Aethopyga saturata</i> | Sch IV | Least Concern |
| 396 | Nectariniidae | Crimson Sunbird | <i>Aethopyga siparaja</i> | Sch IV | Least Concern |
| 397 | Nectariniidae | Fire-tailed Sunbird | <i>Aethopyga ignicauda</i> | Sch IV | Least Concern |
| 398 | Nectariniidae | Little Spiderhunter | <i>Arachnothera longirostra</i> | Sch IV | Least Concern |
| 399 | Nectariniidae | Streaked Spiderhunter | <i>Arachnothera magna</i> | Sch IV | Least Concern |
| 400 | Passeridae | House Sparrow | <i>Passer domesticus</i> | Sch IV | Least Concern |
| 401 | Passeridae | Eurasian Tree Sparrow | <i>Passer montanus</i> | Sch IV | Least Concern |
| 402 | Ploceidae | Baya Weaver | <i>Ploceus philippinus</i> | Sch IV | Least Concern |
| 403 | Estrildidae | Red Avadavat | <i>Amandava amandava</i> | Sch IV | Least Concern |
| 404 | Estrildidae | White-rumped Munia | <i>Lonchura striata</i> | Sch IV | Least Concern |
| 405 | Estrildidae | Chestnut Munia | <i>Lonchura atricapilla</i> | Sch IV | Least Concern |
| 406 | Prunellidae | Alpine Accentor | <i>Prunella collaris</i> | Sch IV | Least Concern |
| 407 | Prunellidae | Rufous-breasted Accentor | <i>Prunella strophciata</i> | Sch IV | Least Concern |

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|-----|--------------|--|--|--------|---------------|
| 408 | Prunellidae | Maroon-backed Accentor Western Yellow Wagtail | <i>Prunella immaculata</i> | Sch IV | Least Concern |
| 409 | Motacillidae | Wagtail | <i>Motacilla flava</i> | Sch IV | Least Concern |
| 410 | Motacillidae | Citrine Wagtail | <i>Motacilla citreola</i> | Sch IV | Least Concern |
| 411 | Motacillidae | Grey Wagtail | <i>Motacilla cinerea</i> | Sch IV | Least Concern |
| 412 | Motacillidae | White Wagtail | <i>Motacilla alba</i> | Sch IV | Least Concern |
| 413 | Motacillidae | Paddyfield Pipit | <i>Anthus rufulus</i> | Sch IV | Least Concern |
| 414 | Motacillidae | Blyth's Pipit | <i>Anthus godlewskii</i> | Sch IV | Least Concern |
| 415 | Motacillidae | Olive-backed Pipit | <i>Anthus hodgsoni</i> | Sch IV | Least Concern |
| 416 | Motacillidae | Rosy Pipit | <i>Anthus roseatus</i> | Sch IV | Least Concern |
| 417 | Elachuridae | Spotted Elachura | <i>Elachura formosa</i> | Sch IV | Least Concern |
| 418 | Fringillidae | Collared Grosbeak | <i>Mycerobas affinis</i> | Sch IV | Least Concern |
| 419 | Fringillidae | Brown Bullfinch | <i>Pyrrhula nipalensis</i> | Sch IV | Least Concern |
| 420 | Fringillidae | Grey-headed Bullfinch | <i>Pyrrhula erythaca</i> <i>Pyrrhoplectes</i> | Sch IV | Least Concern |
| 421 | Fringillidae | Golden-naped Finch Dark-breasted | <i>epauletta</i> | Sch IV | Least Concern |
| 422 | Fringillidae | Rosefinch Plain Mountain | <i>Procarduelis nipalensis</i> | Sch IV | Least Concern |
| 423 | Fringillidae | Finch | <i>Leucosticte nemoricola</i> | Sch IV | Least Concern |
| 424 | Fringillidae | Common Rosefinch | <i>Carpodacus erythrinus</i> | Sch IV | Least Concern |
| 425 | Fringillidae | Scarlet Finch Dark-rumped | <i>Carpodacus sipahi</i> | Sch IV | Least Concern |
| 426 | Fringillidae | Rosefinch | <i>Carpodacus edwardsii</i> | Sch IV | Least Concern |
| 427 | Fringillidae | Crimson-browed Finch Black-headed | <i>Carpodacus</i> <i>subhimachalus</i> | Sch IV | Least Concern |
| 428 | Fringillidae | Greenfinch | <i>Chloris ambigua</i> | Sch IV | Least Concern |
| 429 | Emberizidae | Crested Bunting | <i>Emberiza lathamii</i> | Sch IV | Least Concern |
| 430 | Emberizidae | Tristram's Bunting | <i>Emberiza tristrami</i> | Sch IV | Least Concern |
| 431 | Emberizidae | Little Bunting | <i>Emberiza pusilla</i> | Sch IV | Least Concern |
| 432 | Emberizidae | Chestnut Bunting Black-faced | <i>Emberiza rutila</i> <i>Emberiza</i> | Sch IV | Least Concern |
| 433 | Emberizidae | Bunting | <i>spodocephala</i> | Sch IV | Least Concern |

APPENDIX IV: CHECKLIST OF MAMMALS OF THE DIBANG VALLEY
(Incomplete list)

| | Common name | Scientific name | WLPA 1972 | IUCN | Source |
|----|------------------------|-------------------------------------|---------------------|----------------|---------------|
| 1 | Tiger | <i>Panthera tigris</i> | Schedule I | EN | CT |
| 2 | Clouded leopard | <i>Neofelis nebulosa</i> | Schedule I | VU | CT |
| 3 | Asiatic golden cat | <i>Pardofelis temminckii</i> | Schedule I | NT | CT/DS |
| 4 | Marbled cat | <i>Pardofelis marmorata</i> | Schedule I | NT | CT |
| 5 | Leopard Cat | <i>Prionailurus bengalensis</i> | Schedule I | LC | CT/DS |
| 6 | Asiatic wild dog | <i>Cuon alpinus</i> | Schedule II | EN | CT |
| 7 | Himalayan black bear | <i>Ursus thibetanus</i> | Schedule II | VU | CT |
| 8 | Red panda | <i>Ailurus fulgens</i> | Schedule I | EN | CT |
| 9 | Spotted linsang | <i>Prionodon pardicolor</i> | Schedule I | LC | CT/DS |
| 10 | Masked palm civet | <i>Paguma larvata</i> | Schedule II | LC | CT |
| 11 | Yellow throated marten | <i>Martes flavigula</i> | Schedule II | LC | CT/DS |
| 12 | Yellow bellied weasel | <i>Mustela kathiah</i> | Schedule II | LC | CT |
| 13 | Siberian weasel | <i>Mustela sibirica</i> | Schedule II | LC | CT |
| 14 | Smooth-clawed otter | <i>Lutrogale perspicillata</i> | Schedule II | VU | CT |
| 15 | Mishmi takin | <i>Budorcas taxicolor taxicolor</i> | Schedule I | EN | CT/DS |
| 16 | Himalayan serow | <i>Capricornis sumatraensis</i> | Schedule I | NT | CT/DS |
| 17 | Gongshan muntjac | <i>Muntiacus gongshanensis</i> | Status undetermined | Data deficient | CT/DS |
| 18 | Indian muntjac | <i>Muntiacus muntjak</i> | Schedule III | LC | CT/DS |
| 19 | Wild pig | <i>Sus scrofa cristatus</i> | Schedule III | LC | CT/DS |
| 20 | Red goral | <i>Naemorhedus baileyi</i> | Schedule III | VU | CT/DS |
| 21 | Arunachal macaque | <i>Macaca munzala</i> | Status undetermined | EN | CT |
| 22 | Assamese macaque | <i>Macaca assamensis</i> | Schedule II | NT | CT |
| 23 | Hoary-bellied Squirrel | <i>Callosciurus pygerythrus</i> | Schedule II | LC | CT |

| | | | | | |
|----|---|---------------------------------------|---------------------|----|-----------------------|
| 24 | Himalayan striped squirrel | <i>Tamias mccllellandii</i> | Schedule IV | LC | CT |
| 25 | Himalayan pika | <i>Ochotona himalayana</i> | Status undetermined | LC | CT |
| 26 | Mithun | <i>Bos frontalis</i> | Status undetermined | LC | CT |
| 27 | Himalayan jungle rat | <i>Rattus nitidus</i> | Schedule VI | LC | CT |
| 28 | Himalayan musk deer | <i>Moschus leucogaster</i> | Schedule I | EN | HS |
| 29 | Golden jackal* | <i>Canis aureus</i> | Schedule II | LC | CT |
| 30 | Common palm civet* | <i>Paradoxurus hermaphroditus</i> | Schedule II | LC | CT |
| 31 | Indian sambar* | <i>Rusa unicolor</i> | Schedule III | VU | CT |
| 32 | Rhesus macaque* | <i>Macaca mulatta</i> | Schedule II | LC | CT |
| 33 | Malayan porcupine* | <i>Hystrix brachyura</i> | Schedule II | LC | CT |
| 34 | Asiatic brush tailed porcupine* | <i>Atherurus macrourus</i> | Schedule II | LC | CT |
| 35 | Eastern hoolock gibbon (Mishmi Hills subspecies)* | <i>Hoolock leuconedys mishmiensis</i> | Schedule I | VU | DS |
| 36 | Chinese pangolin | <i>Manis pentadactyla</i> | Schedule I | CR | CT (WII 2019) |
| 37 | Particolored Gliding Squirrel | <i>Hylopetes alboniger</i> | Schedule II | LC | Krishna et al. (2016) |
| 38 | Pallas' squirrel | <i>Callosciurus erythraeus</i> | Schedule IV | LC | DS (WII 2019) |
| 39 | Grey Headed Gliding Squirrel* | <i>Petaurista caniceps</i> | Schedule II | LC | Krishna et al. (2016) |
| 40 | Hodgson's Giant Gliding Squirrel* | <i>Petaurista magnificus</i> | Schedule II | LC | Krishna et al. (2016) |
| 41 | Bhutan Giant Gliding Squirrel* | <i>Petaurista nobilis</i> | Schedule II | LC | Krishna et al. (2016) |
| 42 | Red Giant Gliding Squirrel* | <i>Petaurista petaurista</i> | Schedule II | LC | Krishna et al. (2016) |
| 43 | Mishmi Hill Giant Gliding Squirrel | <i>Petaurista mishmiensis</i> | Schedule II | LC | Krishna et al. (2016) |
| 44 | Yunnan Giant Gliding Squirrel | <i>Petaurista yunanensis</i> | Schedule II | LC | Choudhury (2013a) |
| 45 | Malayan Sun bear* | <i>Helarctos malayanus</i> | Schedule I | VU | CT |

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|----|--|---|-------------|----|----------|
| 46 | Leaf-nosed bat [^] | <i>Hipposideros larvatus leptophyllus</i> | - | LC | ZSI 2006 |
| 47 | Pere David's Vole [^] | <i>Eothenomys melanogaster libonotus</i> | Schedule IV | LC | ZSI 2006 |
| 48 | South China field mouse [^] | <i>Apodemus draco</i> | Schedule V | LC | ZSI 2006 |
| 49 | Millard's Rat [^] | <i>Dacnomys millardi wroughtoni</i> | Schedule V | DD | ZSI 2006 |
| 50 | Ryley's Spiny Mouse [^] | <i>Mus cookii nagarum</i> | Schedule V | LC | ZSI 2006 |
| 51 | Brahma white-bellied rat [^] | <i>Niviventer brahma</i> | Schedule V | LC | ZSI 2006 |
| 52 | Himalayan white-bellied Rat [^] | <i>Niviventer niviventer</i> | Schedule V | LC | ZSI 2006 |
| 53 | Greater Horseshoe Bat [^] | <i>Rhinolophus ferrumequinum tragatus</i> | - | LC | ZSI 2006 |

Sources: CT- Camera trap record; DS – Direct sighting; HS – Hunted specimen collected by Nijhawan (2018) and Nijhawan and Mitapo 2020 (unpublished data)

*Evidence exclusively from Lower Dibang Valley district

[^]Evidence from Dihang Dibang Biosphere Reserve (ZSI 2006)